

METHYLENE BLUE UPTAKE AND INTERMOLECULAR INTERACTIONS IN MICROBIAL CELLS THROUGH FLUORESCENCE-LIFETIME IMAGING MICROSCOPY (FLIM)

Caetano Padial Sabino^{1,2,3}, Maurício da Silva Baptista⁴, Martha Simões Ribeiro¹ and Nilton Lincopan^{2,3}

¹Center for Lasers and Applications, Nuclear, and Energy Research Institute, National Commission for Nuclear Energy, São Paulo, 05508-000, SP, Brazil

²Department of Clinical Analysis, School of Pharmaceutical Sciences, University of São Paulo, São Paulo, SP, Brazil

³Department of Microbiology, Institute for Biomedical Sciences, University of São Paulo, São Paulo 05508-000, SP, Brazil

⁴Department of Biochemistry, Institute of Chemistry, University of São Paulo, University of São Paulo, São Paulo 05508-000, SP, Brazil
caetanosabino@gmail.com

Antimicrobial photodynamic therapy (APDT) is a promising tool to counterattack the emerging treat of drug-resistant pathogens. The technique combines low-intensity monochromatic light with a photosensitizer compound to produce reactive oxygen species (ROS) that can damage virtually any type of biomolecules and lead to rapid cell death. Since some ROS present diffusion-limited reactivity, most cell damage is co-localized with photosensitizer accumulation site. Hence, imaging photosensitizer accumulation and fluorescence lifetime in the nanoscale can bring a great level of information to further understand the ultrastructural cellular damage caused by APDT. In this study, we used a FLIM system capable of single-molecule detection to observe the accumulation and interaction sites of methylene blue (MB), a very broadly-used photosensitizer, in yeast, and Gram-positive and Gram-negative bacterial cells. Our data shows fluorescence lifetime contrast, with nanometric resolution, among different cellular structures such as cell wall, membrane and DNA. The images evidenciate differential MB accumulation in microbial cells and the existence of two different populations of MB molecular species: those interacting mostly with the solvent (short-lived, ~0.8 ns) and those interacting with biomolecules (long-lived, ~2 ns). The short-lived fluorescence predominates in the mucoid capsule of Gram-negative bacteria and cell-wall of yeast and Gram-positive bacteria while long-lived MB fluorescence shows preferential accumulation in DNA-rich sites¹. It is marked in yeast nucleus and exclusively inside bacterial cells. In fact, literature supports that MB intercalation in nucleic acids stabilizes its excited-states leading to increased fluorescence lifetime and efficiency of singlet-oxygen production². Our data brings evidence that this sort of phenomena can be observed by FLIM in the nanoscale and this should bring new insights to the photophysical, photochemical and biological mechanisms of photodynamic therapy.

References

1. Nogueira JJ, Gonzalez L. Molecular dynamics simulations of binding modes between methylene blue and DNA with alternating GC and AT sequences. *Biochemistry* 2014; 53(14): 2391-412.
2. Nogueira JJ, Oppel M, Gonzalez L. Enhancing intersystem crossing in phenothiazinium dyes by intercalation into DNA. *Angew Chem Int Ed Engl* 2015; 54(14): 4375-8.

Acknowledgements: The authors thank all financial support provided by the Brazilian fostering agencies FAPESP, CNPq and CAPES.