



# Advances in Metallic-Based Localized Surface Plasmon Sensors for Enhanced Tropical Disease Detection: A Comprehensive Review

Sajid Farooq<sup>1</sup> · Denise Maria Zezell<sup>1</sup>

Received: 28 September 2023 / Accepted: 23 October 2023 / Published online: 21 November 2023  
© The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2023

## Abstract

Tropical diseases present significant challenges to global health, particularly in resource-limited regions. Early and accurate detection of these diseases is vital for effective management and control. In recent years, metallic-based LSPR sensors have emerged as promising diagnostic tools for sensitive and rapid detection of tropical diseases. This comprehensive review aims to provide an in-depth analysis of the current state of research on metallic-based LSPR sensors for the detection of various tropical diseases. In this study, we focused on the connection between neglected tropical diseases (NTDs) and its risk using metallic-based LSPR sensors to identify potential inflammatory biomarkers. We conducted a literature search using PubMed, Web of Science, and Google Scholar. Only published materials written in English were considered, resulting in the identification of ~ 220 articles. After a comprehensive evaluation, we selected 35 relevant ones. Our analysis revealed 35 links to neglected tropical diseases, providing valuable insights into their relationship using metallic-based LSPR sensors. Moreover, we explore the potential of metallic-based LSPR sensors in point-of-care testing and their integration with emerging technologies such as microfluidics and smartphone-based diagnostics. This review underscores the need for continued research efforts to develop affordable, sensitive, and user-friendly metallic-based LSPR sensors for early detection and surveillance of tropical diseases.

**Keywords** Metallic nanoparticles · Tropical diseases · Optical sensors · Sensitivity · Limit of detection · Localized field

## Introduction

The optical properties of plasmonic nanoparticles (NPs) and their surface-plasmon resonances have garnered potential interest due to their unique characteristics. Unlike plasmons supported on bulk metal surfaces, NP plasmons present quantized electron oscillations confined to nanoscale volumes. This confinement enables manipulation of light-matter interactions while surpassing the limitations imposed by diffraction [1–3]. These properties have paved the way for diverse applications in the domain of optics [4], electronics [5], sensing [6], photonics [7], and medical diagnostics [8]. For instance, the advancement of miniaturized optical and electronic nanodevices, nanosensors, and nanophotonic circuits has been facilitated by leveraging the distinctive optical

characteristics of nanoscale metals. Hence, the integration of plasmonic systems in medical diagnostics and therapeutics holds promise for improved bio-medical imaging [9], targeted drug delivery [10], and biosensing [11]. Recent theoretical advancements in NP-based plasmonics have led to valuable insights for the design and understanding of optical nanostructures. While Mie's theory of light scattering and absorption by gold (Au) nanospheres dates back over a century, modern computational frameworks and theoretical developments have enabled for the exploitation of plasmon resonances in more intricate NP architectures and assemblies [12]. In particular, plasmon hybridization theory has emerged as a powerful theoretical framework that draws parallels between plasmons in metallic nanoparticle assemblies and the behavior of electrons in quantum molecular orbitals [13, 14]. This theory predicts that plasmons on adjacent metallic nanostructures interact, mix, and hybridize, analogous to the behavior of electronic wave functions in simple atomic and molecular orbitals. The insights gained from plasmon hybridization theory hold promise for the rational assembly of plasmonic NPs into well-defined structures such

✉ Sajid Farooq  
sajiddahar@gmail.com

<sup>1</sup> Center for Lasers and Applications, Nuclear and Energy Research Institute, IPEN—CNEN, Sao Paulo 05508-000, Brazil

as plasmonic molecules, polymers, and crystals. By tailoring the arrangement and properties of these plasmonic nanostructures, it becomes possible to engineer materials with customizable optical properties, opening up new horizons for advanced optical and photonic applications.

Conductive nanostructures, in particular, exhibit strong interactions with light, leading to a collective oscillations of conduction electrons, termed surface plasmon (SP) [15–18]. When the coherent collective oscillations of electrons occur under resonance conditions, it is referred as Localized Surface Plasmon Resonance (LSPR), which can outcome in highly efficient light absorption or scattering and robust field enhancement [3, 19–21]. The plasmon resonance phenomenon heavily depends on the composition [20, 22, 23], size [20, 24], and shape [3, 25, 26] of the nanostructures, as well as the surrounding environment [27–29]. Various types of materials, in a different configuration, can assist plasmons, including colloid dispersion's isotropic [30–35] and anisotropic [36–39] and anisotropic metallic NPs [40–44], metal oxide NPs [45–48], quantum dots [49–52], organized metallic nanostructures [53–56], and two-dimensional materials such as graphene [57–60].

However, nanomaterials composed of copper (Cu) [61–64], silver (Ag) [65–67], and gold (Au) [68, 68–71] have garnered special attention due to their LSPR occurring within the optical range of the electromagnetic spectrum. These materials exhibit unique optical plasmonic properties that may be harnessed for a wide range of applications.

Recently, LSPR sensors emerged as outstanding candidates for sensitive, facile, and rapid detection of tropical diseases [72–76]. LSPR sensors exploit the unique optical features of metal-based NPS, typically Au, Ag, or Cu, to detect molecular interactions at the nanoscale. By leveraging the principles of surface plasmon or localized plasmon resonances, such types of sensors enable label-free [77–79] and real-time detection [80–83] of disease-specific biomarkers with exceptional selectivity and sensitivity. The domain of LSPR sensors for tropical disease detection has witnessed significant advancements, with researchers exploring various strategies to improve detection accuracy, miniaturize sensor platforms, and integrate them with portable and point-of-care devices [72, 84–86]. These advancements have the potential to revolutionize the diagnosis and surveillance of tropical diseases, particularly in resource-constrained settings where access to centralized laboratory facilities is limited.

This review focuses on providing a detailed analysis of the current state-of-research on LSPR sensors for the detection of tropical diseases. It will delve into the underlying principles of plasmonic sensing, elucidate the advantages and challenges associated with these sensors, and underscore recent progress in their applications for the recognition of specific tropical diseases, including malaria, dengue fever, chikungunya, and others. Furthermore, this review will

explore the potential of LSPR sensors in point-of-care testing and their integration with emerging technologies such as microfluidics and smartphone-based diagnostics (Fig. 1). By critically examining the progress made in this field, this review underscores the importance of ongoing research efforts in developing affordable, sensitive, and user-friendly plasmonic sensors for early detection and surveillance of tropical diseases. Ultimately, the widespread adoption of LSPR sensors has the potential to enhance disease control strategies, reduce morbidity and mortality rates, and contribute to the global effort to combat tropical diseases.

## Tropical Disease

The neglected tropical diseases (NTDs) are a group of infectious diseases, predominantly affecting populations in tropical and subtropical regions. These diseases, including Leptospirosis, Chagas Disease, Dengue Fever, and others, pose significant health challenges in these areas [87–91]. NTDs are caused by various pathogens and are closely linked to environmental factors such as climate, sanitation, and disease vectors [92–94]. Socioeconomic factors like poverty, limited healthcare infrastructure, and lack of access to clean water further exacerbate the burden of NTDs, perpetuating a cycle of poverty and ill health in affected communities [95–98].

A plethora of literature highlights ongoing research efforts to improve NTD diagnosis and management. These efforts involve innovative diagnostic tools like electrochemical biosensors [99], CRISPR-based biosensors [100], and surface plasmon resonance (SPR) biosensors [101]. Additionally, researchers are exploring remote sensing technologies and terahertz (THz) sensors for disease mapping and detection [102, 103]. These studies provide valuable insights and potential solutions for the detection and understanding of NTDs.

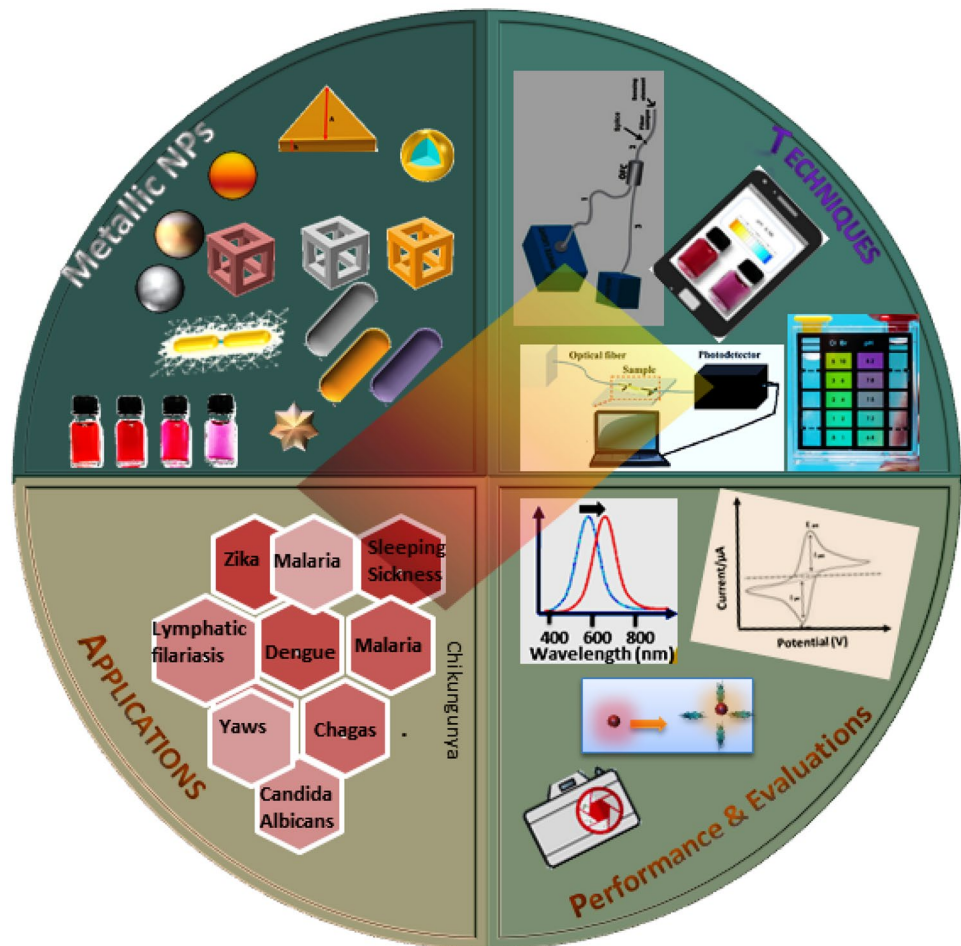
To enhance the development of sensitive and reliable sensing platforms for NTDs, it is important to meticulously address various factors including careful material selection, surface functionalization, and consideration of factors like cross-sensitivity and selectivity among multiple analytes. Metallic-based LSPR sensors are highlighted as promising tools due to their cost-effectiveness, user-friendliness, sensitivity, and portability [104–106]. Overall, LSPR-based sensors underscore the significance of ongoing scientific research in addressing the complex challenges posed by NTDs.

## Methods

### Study Selection

This comprehensive review involved accessing and evaluating articles related to metallic-based sensors for the analysis

**Fig. 1** Graphical representation of the metallic NPs-based on shape, size, materials composition, and spectroscopic techniques for performance evaluation to identify neglected tropical disease



of Neglected Tropical Diseases (NTDs). In 2023, we conducted three independent searches on electronic databases, including Web of Science, Google Scholar, and PubMed, using the keywords “LSPR sensors,” “tropical disease,” and “metallic nano-structures” to identify relevant studies for further analysis.

### Data Extraction

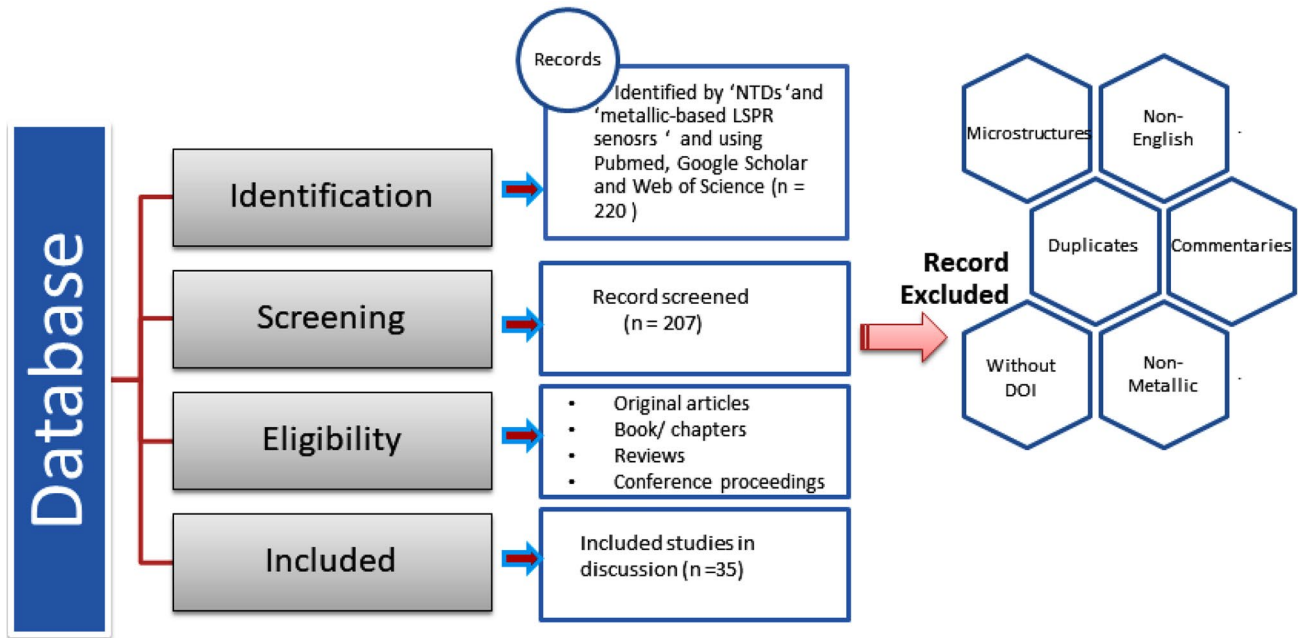
We further narrowed down our selection to articles published within the last decade. The initial list underwent a preliminary screening, focusing on experimental studies. We specifically looked for original articles written in English while excluding review articles, conference abstracts, letters, book chapters, case studies, reports, and editorials, including method and protocol articles. To ensure thoroughness, two independent investigators evaluated the titles and abstracts for eligibility. Articles that met these criteria had their conclusions and texts reviewed to determine eligibility. We selected articles for our review if they addressed the signaling metallic-based NPs, sensitivity, Limit of Detection (LoD), and their relation to Neglected Tropical Diseases

(NTDs), regardless of their methodological approaches. A visual representation of our inclusion process can be found in Fig. 2.

## Principles of Plasmonic Sensing

### Surface Plasmon Resonance

Ritchie and colleagues introduced the term “SPP” while studying the angle-energy distribution involved in the energy transfer process between an incident electron beam and conduction electrons within a thick metallic foil [107]. Their investigation revealed an intriguing pattern: the energy loss exhibited an inverse relationship with the film thickness. This observation showed at the potential involvement of Surface Plasmon Polaritons (SPPs) in the energy transfer process. Notably, materials like Au, Ag, and Cu have gained significant attention for studying SPPs due to their distinctive dielectric properties—characterized by highly negative real parts and relatively low positive imaginary parts (Figs. 3 and 4) [108]. A distinguishing

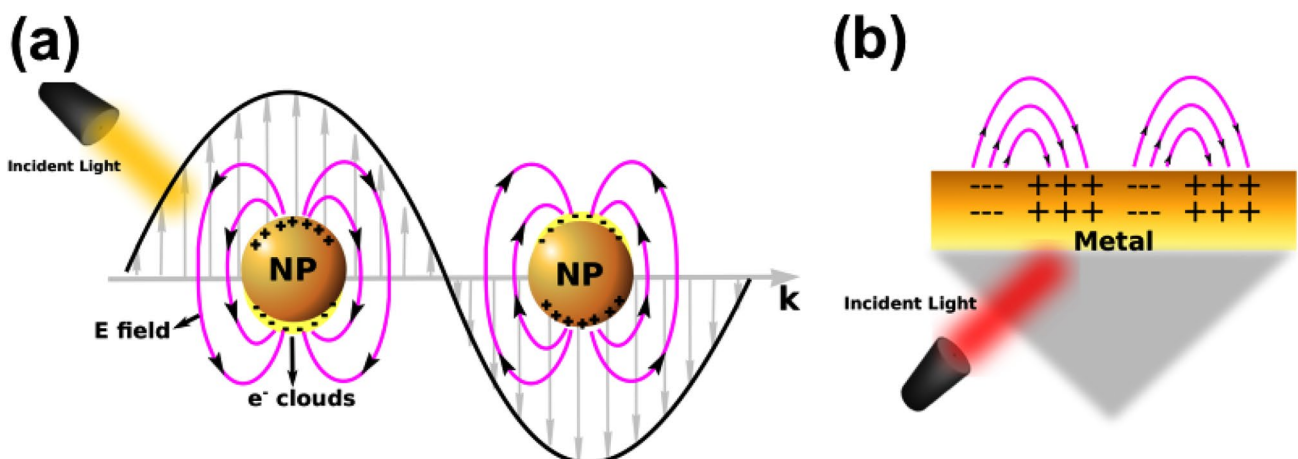


**Fig. 2** Flowchart depicting the process of article identification across three independent electronic databases (Web of Science, PubMed, and Google scholar), leading to the inclusion of a total of 35 articles in the final discussion

factor setting noble metals apart from other metallic counterparts lies in their imaginary dielectric function components. As depicted in Fig. 4a, the imaginary component for Ag remains below 2 within the visible spectrum wavelength range. Similarly, Au displays a distinct minimum of around 700 nm wavelength. Particularly, intriguing is Ag's behavior, where the imaginary component stays significantly below 1 for wavelengths below 400 nm. In Fig. 4b, the red curve representing the real part of the dielectric function for Ag follows analogous patterns. It

maintains a consistently negative value, dropping below  $-20$  for wavelengths around 600 nm. This feature renders such materials exceptionally appropriate for SP coupling. Moreover, noble metals exhibit SPP coupling frequencies within the visible region of the electromagnetic spectrum, simplifying their practical application [107, 108].

Multiple techniques are available to generate SPPs in metallic thin films. These methods include using tightly focused light beams [109], employing diffraction gratings [110], or using prisms to ensure precise incidence angles



**Fig. 3** Schematic diagram of the plasmonic resonance-based phenomena in two different configurations: **a** SPR behavior and **b** LSPR phenomenon

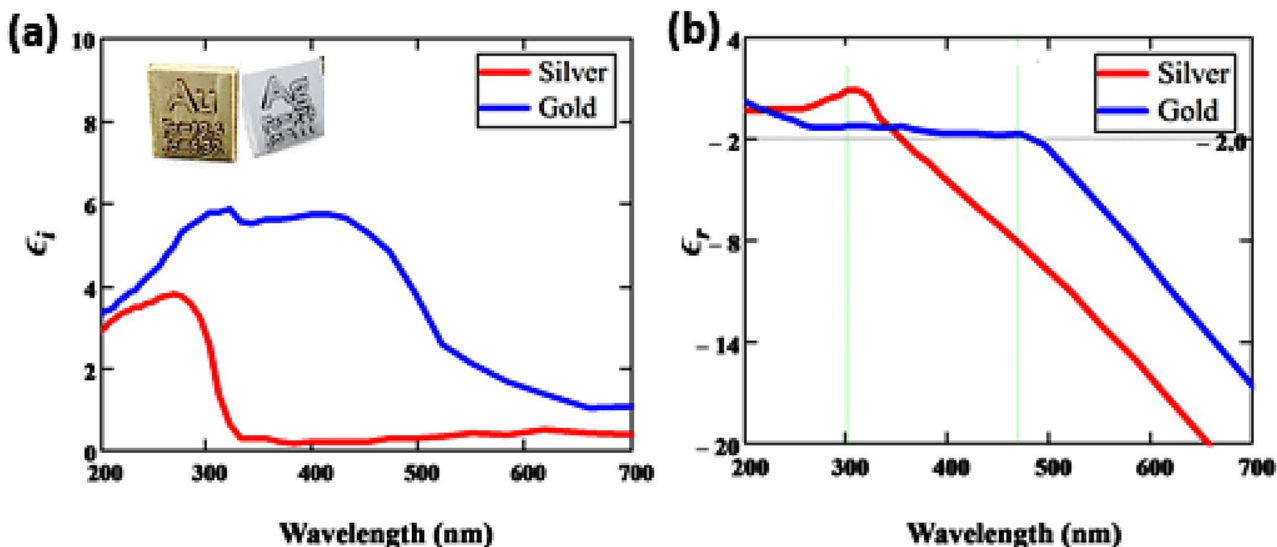


Fig. 4 a Imaginary and b real parts of the dielectric function of Ag and Au [44]

[111]. Regardless of the selected approach, a crucial prerequisite for successful SPP excitation is the alignment of the incident light beam’s wavevector ( $k$ ) with the SPP wavevector ( $k_{sp}$ ) to assist effective coupling. Prism-based strategies can be further categorized into distinct setups, commonly known as the Kretschmann and Otto configurations. The Kretschmann configuration [112] involves positioning the metal film in direct contact with the prism’s surface. Hence, the incident light beam is directed at an angle that enables a harmonious alignment of  $k$  and  $k_{sp}$ , promoting coupling between them. On the other hand, the Otto configuration [113] shares similar conditions for exciting SPPs. However, in this case, the SPP formation occurs within a gap—either filled with air or a dielectric solution—placed between the metallic film and the prism.

In both of these prism-coupling set-up configurations, the SPP is excited using a monochromatic beam, commonly employing a red laser, which is specifically well-suited for Au films. This selection of laser wavelength ensures that the value of  $k$  is defined as follows:

$$k = (\omega_0/c)\eta_0 \tag{1}$$

Here,  $\omega_0$  represents the frequency of the incident light,  $\eta_0$  stands for the refractive index of the prism, and  $c$  signifies the velocity of light in a vacuum. Then, SPP wavevector is given by [114]:

$$k_{sp} = (\omega_0/c) \left[ \frac{(\epsilon_1 \eta_2^2)}{(\epsilon_1 + \eta_2^2)} \right]^{1/2} \tag{2}$$

In this context,  $\epsilon_1$  denotes the dielectric function of the metal,  $\eta_2 = \sqrt{\epsilon_2}$  represents the refractive index (RI) of the

medium situated adjacent to the metallic film (generally the analyte in sensor applications), and  $\epsilon_2$  relates to its respective dielectric function. The coupling phenomenon is obtained through precise adjustment of the angle of incidence of light on the metal surface, denoted as  $\theta$ , ensuring that the projection of the wavevector of light onto the surface aligns with the plasmon wavevector:  $k \sin \theta = k_{sp}$ .

### Localized Surface Plasmon Resonance

Mie [12] introduced the first theoretical analysis of the LSPR phenomenon. By solving Maxwell’s equations for an electromagnetic wave that interacts with a sphere significantly smaller than the wavelength of incident light ( $2r \ll \lambda$ , where  $r$  represents the sphere’s radius), Mie derived an equation to determine the particle’s polarizability and subsequently, for the particle’s extinction cross-section [12, 115]. The polarizability is mathematically expressed as follows:

$$\alpha = 4\pi r^3 \frac{\epsilon_1 - \epsilon_2}{\epsilon_1 + 2\epsilon_2} \tag{3}$$

where  $\epsilon_1$  corresponds to the particle’s dielectric function. It becomes evident that  $\alpha$  undergoes a phenomenon of resonant enhancement when the real part of  $\epsilon_1$  equals  $-2\epsilon_2$ , signifying the approach towards a minimum value of  $|\epsilon_1 + 2\epsilon_2|$ . This relationship is commonly referred to as the Fröhlich condition [115–117]. The extinction cross-section of a NP is determined by the combined effects of its absorption ( $\sigma_{abs}$ ) and scattering ( $\sigma_{sca}$ ) cross-sections. These characteristics rely on the particle’s polarizability and can be expressed as follows:

$$\sigma_{sca} = \frac{k^4}{6\pi} |\alpha|^2, \quad (4)$$

$$\sigma_{abs} = k \operatorname{Im}[\alpha]. \quad (5)$$

It's evident that the  $\sigma_{abs}$  and  $\sigma_{sca}$  exhibit distinct dependencies on the particle's radius. Consequently, the particle size plays a pivotal role in shaping its LSPR.

## Factors Affecting LSPR

In the following sections, we will briefly discuss the LSPR dependence on the material composition, size, and shape.

### Material-Dependent LSPR

Theoretical realization of LSPR extends to a variety of materials including metals, semiconductors, and alloys, contingent upon fulfilling the Fröhlich condition and featuring a real part of the dielectric function ( $\operatorname{Re}[\epsilon_1]$ ) less than zero due to the presence of conduction electrons. Notably, in the realm of LSPR applications, copper (Cu), gold (Au), and silver (Ag) emerge as the most prominent metals, presenting LSPR spectral peaks within the visible spectrum. Furthermore, Al introduces a plasmon resonance in the ultraviolet region where several organic molecules exhibit light absorption [118]. Further elements like sodium (Na), lithium (Li), and gallium (Ga) also meet the Fröhlich criterion, unveiling plasmon resonance within the UV-Visible spectrum [119–122]. Nevertheless, the susceptibility of these metals to oxidation and their reactivity to the environment restricts their use as sensor platforms.

In a broad context, materials satisfying the Fröhlich criterion demonstrate a negative real part ( $\epsilon_r$ ) and a slightly positive imaginary part ( $\epsilon_i$ ) within the dielectric function. In the specific case of a metallic nanosphere situated in a vacuum, the Fröhlich criterion aligns with ( $\epsilon_r$ ) approximately equal to  $-2$ . This criterion is attained at wavelengths of about 350 nm for Ag, and approximately 490 nm for Au, as indicated in 4.

In the context of LSPR sensing, the preference lies with an imaginary part of the dielectric function featuring modest positive values, indicative of low electron energy loss, thus resulting in narrow LSPR linewidths. Specifically, at the point of LSPR resonance, the imaginary parts of the dielectric function are 3.81 for Au (490 nm) and 0.28 for Ag (at 350 nm), as illustrated in (Fig. 4).

### Size-Dependent LSPR

The interaction between light and metallic NPs is strongly affected by the size of these structures. Reducing the size of metallic NPs amplifies the dipole contribution to the LSPR

spectra. Conversely, increasing the radius of NPs introduces the potential for higher order modes, such as quadrupoles, in the case of Ag NPs, to significantly impact the particle's extinction cross-section. Notably, both  $\sigma_{sca}$  and  $\sigma_{abs}$  exhibit size-dependent behavior. Within a quasi-static approximation, the absorption cross-section of a spherical NP scales with the third power of the NP radius, while the scattering cross section relies on the sixth power of the NP's radius. Consequently, for larger metallic NPs (>50 nm), the light-NP interaction is primarily governed by scattering. In this scenario, enhancing the size of metallic NPs leads to augmented scattering cross sections [124, 125]. However, this enlargement is accompanied by a rise in radiation damping, resulting in spectral broadening and shifts [126, 127].

In the case of particles with small radii ( $\leq 15$  nm), dipolar absorption dominates the extinction spectra. This implies that scattering, being a radiative phenomenon, is generally of lesser significance. As shown in Fig. 5, the broadening of LSPR spectrum increase as a function of size as well as the shifting of the LSPR spectra. However, Farooq et al. observed that spectral changes are more dominant for Ag NPs than for Au nanostructures [20]. Furthermore, the spectral resonance peak shift and sensitivity of different sizes of NPs can be seen in Table 1.

### Shape-Dependent LSPR

Tuning the morphology of metallic NPs enables to control and fine-tune their LSPR characteristics, enabling the manipulation of plasmon peaks across a range of spectral regions. Moreover, NPs with sharper tips are contributed to exhibit LSPR at longer wavelengths compared to spherical ones. For instance, as shown in Fig. 5b. in the case of Ag NPs, nanospheres exhibit LSPR peak in the blue region, while Ag nanocubes depict an extinction peak at a green wavelength. With even more sharper tips, such as in nanotriangle structures, there's a clear shift towards longer wavelengths, resulting in a red-shifted plasmonic peak when compared to cubic nanostructures. Additionally, Chen et al. conducted a study involving Au NPs with various shapes, including nanospheres, nanorods, nanocubes, nanobranches, and nanobipyramids. The study revealed variations spanning from the visible to the near-infrared region as a function of shape [128, 129]. The plasmonic behavior and RI based sensitivity as a function of shape can be seen in Table 2.

### Materials-Dependent LSPR

LSPR can be theoretically achieved in various metals, semiconductors, or alloys by fulfilling the Fröhlich condition. Among metals, Au and Ag are exclusively utilized for LSPR applications due to their negative real part and positive imaginary part of the dielectric functions. Moreover,

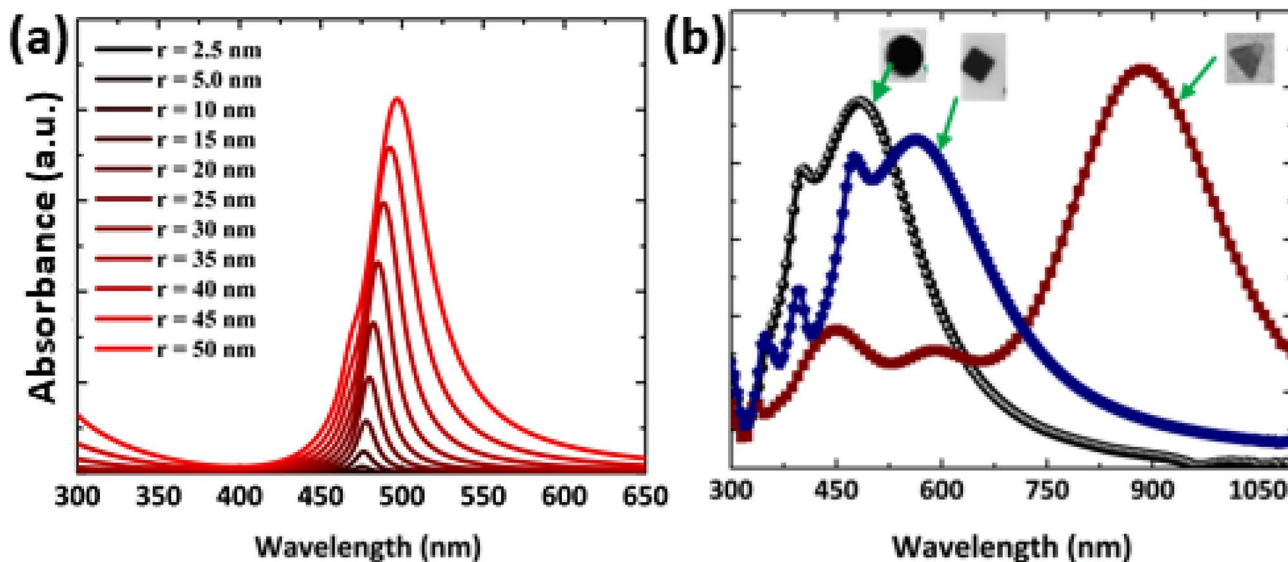


Fig. 5 a The optical properties of metallic NPs: a size dependence [5] and b shape dependence LSPR [123]

aluminium (Al) shows plasmon resonance in the ultraviolet region, which is impactful for light absorption by organic molecules. Copper (Cu) is another optical plasmonic metal that shows LSPR resonance, similar to Ag and Au NPs, with a narrow and intense peak. Other metals such as sodium (Na), Lithium (Li), and Gallium (Ga) may also follow the Fröhlich criterion, depicting plasmon resonance in the UV-Visible spectrum. However, their use in experiments is challenging due to their high reactivity and susceptibility to oxidation compared to Au or Ag. For Au and Ag, the LSPR can be tuned to the UV-Vis or near-IR spectrum. As shown in Fig. 4, the real parts of the dielectric functions for Au and Ag demonstrate negative values of  $\epsilon_r$ , fulfilling the Fröhlich condition for a NP in air.

### Localized Field Enhancement

As Fröhlich condition is satisfied, it leads to the enhancement of the localized field both inside and outside of NP. This enhancement becomes measurable through the evaluation of the electric field distributions within ( $E_{in}$ ) and outside ( $E_{out}$ ) of a NP. [130]:

$$E_{in} = \frac{3\epsilon_2}{\epsilon_1 + 2\epsilon_2} \times E_0 \tag{6}$$

$$E_{out} = E_0 + \frac{3\mathbf{n}(\mathbf{n} \cdot \mathbf{p}) - \mathbf{p}}{\epsilon_1 + 2\epsilon_2} \times E_0 \frac{1}{r^3} \tag{7}$$

where  $\mathbf{n}$  and  $\mathbf{p}$  denote the unit vector normal to the surface and dipole moment, respectively.

The enhancement of the localized field ( $EF_F$ ) near the surface of the NP is given as follows:

$$EF_F = \frac{|E_{max}|}{|E_0|} \tag{8}$$

where  $E_0$  represents the incident field and  $E_{max}$  attributes the maximum field in the vicinity of NP’s surface.

Increase localized field effects are more associated in non-spherical structures when they resonate plasmonically. Figure 6 represents the field enhancement across different shapes of Ag NPs – nanospheres, nanocubes, and nanotriangles – immersed in a  $H_2O$  environment. Remarkably improved  $EF_F$  values are observed, especially near the tips

Table 1 The size-dependent sensitivity and figures of merit for LSPR platform

Au nano spheres	Matrix type	LSPR peak (nm)	RIS (nmRIU <sup>-1</sup> )	FoM (nmRIU <sup>-2</sup> )	References
5 nm	Single	520	~ 44	0.5	[136]
15 nm	Ensemble	527	44	0.6	[137]
30 nm	Ensemble	530	60	-	[138]
60 nm	Ensemble	535	90	1.5	[139]
100 nm	Ensemble	552	185width at half maximum (FWHM) of the LSPR	1.9	[24]

**Table 2** Diverse shapes of metallic-based LSPR sensors with their sensitivity and FoM

Nanostructure	Types	<i>RIS</i> (nm)	FoM (nm RIU <sup>-1</sup> )	<i>RIS</i> × FoM (nm RIU <sup>-2</sup> )	References
Au shell	Single	381	5.4	2057.4	[140]
Au star	Single	238	1.9	452.2	[144]
Au rattle	Single	199	3.8	756.2	[145]
Au crescent	Ensemble	596	2.4	1430.4	[146]
Au bipyramid	Ensemble	353	-	1584.0	[147]
Au dimeric NR	Single	544	12.4	6745.6	[40]

of these nanostructures. Consequently, non-spherical metallic NP possess considerable appeal for a range of plasmonic applications, including Surface-Enhanced Raman Spectroscopy (SERS) [131], fluorescence sensing [44], and imaging [132] (Fig. 7).

## LSPR Sensing Mechanisms

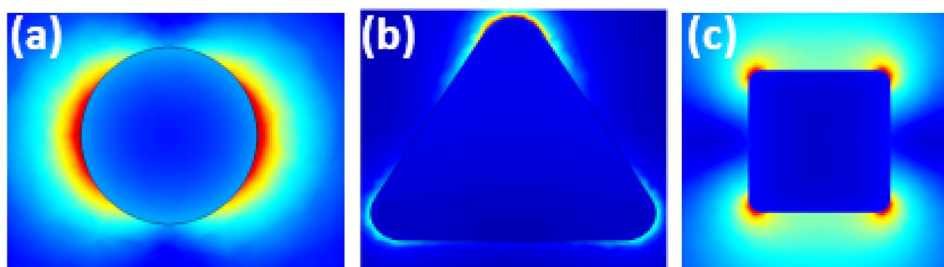
### Refractive Index Based Sensing

The relationship between the LSPR extinction spectrum and the permittivity of the surrounding medium has been a subject of thorough investigation for sensing applications. When the dielectric permittivity of the surrounding medium increases, it induces a red-shift in the plasmon resonance peak [133]. Evaluating the effectiveness of an LSPR sensor often involves the consideration of bulk sensitivity (*RIS*), quantified as the change in LSPR peak wavelength per unit alteration in the refractive index (RIU) [134, 135].

$$RIS = \frac{\Delta\lambda_{LSPR}}{\Delta n_m} \quad (9)$$

where  $\Delta\lambda_{LSPR}$  represents the shift in the wavelength of the LSPR peak, and  $\Delta n_m$  signifies the alteration in the refractive index of the surrounding medium.

**Fig. 6** Localized field enhancement of diverse shapes of Ag NPs: **a** sphere, **b** triangle, and **c** cube [123]



## Figure of Merit

Precise shifts within the resonance peak can sometimes be challenging to discern, especially amidst a broad LSPR spectrum. For a comprehensive assessment of the sensor's efficacy, we introduce another parameter, the figure of merit (FoM). This value represents the ratio of bulk sensitivity (*RIS*) to the full width at half maximum (FWHM) of the LSPR peak (FoM = *RIS*/FWHM). Consequently, an optimal sensor must fulfill both aspects: exhibit substantial bulk sensitivity while maintaining a narrow LSPR spectrum. This criterion finds its expression in the form of *RIS*×*FoM*.

$$FoM = \frac{RIS}{FWHM} \quad (10)$$

where FWHM represents full-width half maximum of the LSPR spectrum.

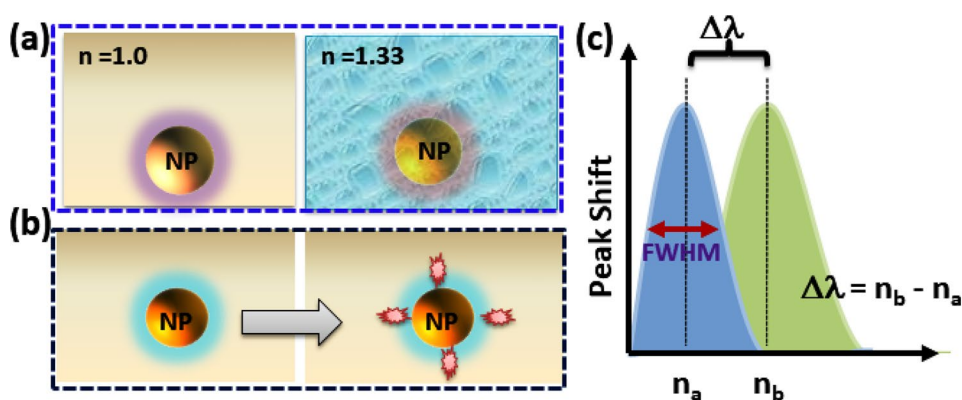
A successful plasmonic sensor demands to simultaneously meeting two essential criteria: achieving high sensitivity while maintaining a distinct and narrow LSPR spectrum. Complex plasmonic structures are frequently explored for sensing purposes due to their ability to provide elevated sensitivity values. However, this pursuit of sensitivity might not consider the broader nature of the resulting LSPR peak.

Consider, for example, LSPR sensors that rely on intricate nanoparticle shapes like nanostars, nanorices, nanorods, and nanocrescents. While these sensors can demonstrate remarkable sensitivity, they might exhibit lower FoM values due to their inherently broader spectral responses. To address this trade-off, Farooq et al. [140] introduced a novel figure of merit (*RIS*×*FoM*) that presents a comprehensive assessment of LSPR sensor performance by considering both sensitivity and spectral width as key factors.

## Molecular Sensing

The LSPR sensor exhibits several advantageous features, making it well-suited for biomedical and biological assays. One of its key strengths is its label-free nature, enabling the detection of molecular attachments to the surface of NPs (NPs). The sensor's sensitivity is enhanced as the

**Fig. 7** A schematic diagram of LSPR sensing mechanism: **a** refractive index based sensing (RIS), **b** molecular sensing phenomenon, and **c** figure of merit



volume of analyte increases [141]. A fundamental phenomenon underlying LSPR-based molecular sensing includes self-assembled monolayer (SAM) on the surface of the NP [142]. Campbell and colleagues developed a mathematical model that elucidates plasmon resonance signals stemming from adsorbed molecular layers on nanostructures [143]. This model is explored as a valuable tool for gauging adlayer thickness on plasmonic structures, and it’s relevant to both SPR and LSPR techniques. The framework of this model revolves around the interplay between the intensity of the electric field (*I*) and the distance (*z*) from the surface. The shift in the LSPR peak due to the adsorption of a molecular monolayer onto a metallic NP surface can be effectively explained by Campbell’s model [143]:

$$\Delta\lambda = RIS(n_{ads} - n_m)(1 - e^{-2d/l_d}) \tag{11}$$

where *l<sub>d</sub>* represents the electromagnetic field decay length around the NP, *d* associates to the thickness of the adsorbate layer, while *n<sub>ads</sub>* and *n<sub>m</sub>* denote the refractive indices of the adsorbate layer and the surrounding medium, respectively.

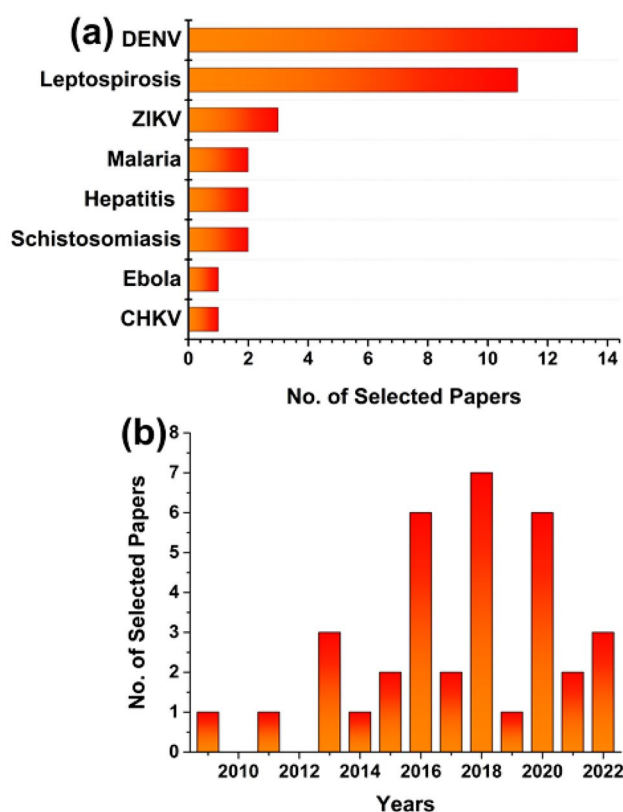
### Results and Discussion

In this comprehensive review, our initial search identified a total of 250 articles from various databases, including Google Scholar, PubMed, and Web of Science (173) (see Fig. 2). After meticulously eliminating duplicate records, both manually and by employing the Bibliography Library, we were left with 220 unique articles. To ensure the relevance and specificity of our review, we excluded articles falling into several categories, including review papers, metallic nanoparticles without NTD, QDs studies, and studies focusing solely on SPR unrelated to NTDs. Additionally, literature that did not encompass tropical diseases was also removed from consideration. As a result of this rigorous screening process, we ultimately included 35

articles in our comprehensive review following a comprehensive full-text assessment.

Among the selected literature, the majority of research focused on NTDs such as DENV, ZIKV, Leishmaniasis, and Schistosomiasis in the context of tropical diseases. Interestingly, there were limited studies specifically addressing metallic-based LSPR sensors unique to NTDs, likely due to the challenges (stability, user friendly, etc.) of metallic NPs with various other medical conditions. To provide a clear description of the distribution of selected articles, Fig. 8a offers a topic-wise classification. We also categorized these articles based on their publication year (Fig. 8b), allowing us to prioritize recent research and technological advancements. Notably, a significant proportion of the selected articles were selected before 2023, which can be attributed to the heightened demand for metallic-based sensors for NTDs. These innovations were crucial for managing tropical diseases, especially those at higher risk, in the comfort of their homes.

Conventional methods of tropical disease detection, involving microscopy, serological assays, and molecular techniques, have been foundational in public health management. However, these methods come with inherent limitations. Microscopy, while widely used, is labor-intensive and may lack the sensitivity required for early disease detection. Additionally, it relies on skilled personnel and can be time-consuming, delaying timely interventions [180, 181]. On the other hand, serological assays can provide false negatives, particularly in the case of low antibody levels, and may not discriminate between active as well as past infections. Such methods can be resource-intensive, necessitating expensive laboratory infrastructure, reagents, and skilled technicians [182, 183]. In the context of conventional studies, the novel diagnostic techniques introduced are not without their own set of limitations. CRISPR-based biosensors, while highly specific, can be complex to design and may demand optimization for diverse diseases. Additionally, these methods may not always offer the desired sensitivity for early disease detection [184, 185]. Surface plasmon resonance (SPR)



**Fig. 8** The selected papers as a function of classification according to a research topic regarding NTDs and **b** according to year of publication

biosensors and electrochemical biosensors, while promising, can be expensive to implement, and their success relies on careful material selection and functionalization [186]. Furthermore, the cost of these technologies can be a hindrance to exclusive adoption in resource-limited settings [187]. Overall, while conventional methods have been employed as the foundation for NTDs detection, they suffer from limitations attributed to sensitivity, specificity, cost, and infrastructure requirements. Conventional studies, while inducing novel diagnostic methods, can limit their applications due to complexity, cost, and precision. The current research in this domain aims to provide a balance between these techniques to enhance disease sensitivity and healthcare results in regions affected by NTDs [188–190].

Plasmonic sensing platforms encompass a wide range of materials including noble metals, transition metallic nitrides, metallic chalcogenides, semiconductor nanocrystals, and metallic oxides [191]. Furthermore, these platforms can be extended to incorporate intricate multicomponent structures such as alloys and core-shell/nanoshell configurations [23, 192]. The LSPR phenomenon exhibited by these materials is finely tuned by factors such as chemical composition, shape, aspect ratio, and size, as aforementioned [193]. However, it's important to note that each material comes with its own

inherent advantages and limitations, both of which hinge on the precise optimization of synthetic conditions. Achieving these optimal conditions is far from a straightforward endeavor, as they can substantially vary due to a multitude of factors extensively explored elsewhere [192, 194, 195]. These factors encompass aspects like the choice of reducing and stabilizing agents, temperature control, deposition substrate, and more. In essence, navigating the path to achieving the best possible conditions is a complex undertaking that requires careful consideration and adjustment across a diverse array of variables.

These platforms offer remarkable versatility, capable of adopting a plethora of architectural designs, all without the need for specific incidence angles, high-intensity light sources, or intricate detection systems. For instance, plasmonic NPs find utility in various formats such as suspensions [196, 197], depositions onto solid substrates like glass, semiconductors [198, 199], metallic films, polymers [200], paper [201, 202], and even fiber optics [203]. The process of illumination and detection can be executed using conventional UV-Vis spectrophotometers, harnessed through dark-field microscopy [204–206], achieved via a simple LED and a cell phone camera [201], or even through direct visual inspection [207]. In fact, these platforms deliver reliable outcomes swiftly and cost-effectively, not only within the controlled confines of a laboratory setting but also in field analyses and locations with restricted access. This adaptability and accessibility underline their significance in numerous practical applications.

The simplest LSPR platform entails utilizing suspended NPs due to its straightforward handling, enabling measurements using a conventional spectrophotometer or even unassisted visual observation [207]. A significant portion of visually interpretable LSPR colorimetric sensing platforms revolves around the aggregation of NPs, a mechanism that facilitates substantial peak shifts. Numerous research endeavors have tackled significant subjects concerning tropical diseases and diagnostic techniques rooted in the LSPR phenomenon. Kim and colleagues engineered a colorimetric rapid diagnostic test (RDT) kit with the ability to detect Zika virus (ZIKV) [208]. This innovative kit employs monoclonal antibodies in conjunction with non-structural protein-1 (NS1) conjugated to Au NPs. Further, the authors revealed that the Zika RDT presented high analytical sensitivity, with 100% identical results compared to ELISA and PCR. Additionally, the kit demonstrated remarkable selectivity, as it did not cross-react with other infected sera, involving DENV, yellow fever, and hepatitis-C virus. Similarly, Brangel and collaborators pioneered a point-of-care test employing an immunochromatographic strip and smartphone reader, which effectively detects Ebola-specific antibodies in human survivors with exceptional sensitivity and specificity [160]. The sensor also involved multiplex

tests for detecting antibodies against multiple viral species, aiding in cross-reactive immunity assessment. The simple, fast, and portable system showed key potential for diagnostic procedures, therapeutic evaluation, and vaccine development in Ebola-affected regions like Uganda. In a different study, Sattarahmady et al. developed a novel diagnostic method for *Leishmania major* using Au NPs and a specific DNA probe [152]. The approach depends on the dispersion of Au NPs probe conjugates based on the presence of a complementary DNA-sequence, leading a sensitive shift in UV-Visible spectra as well as solution color. The method proved efficient in accurately detecting *Leishmania* NTD from other non-*Leishmania* diseases, with a detection limit of  $7.0 \text{ pg } \mu\text{mL}^{-1}$ . Furthermore, the PCR-free assay successfully identified genomic-DNA from, both, clinical samples and *Leishmania major*. This new method offers great promise for improved and efficient diagnosis of *Leishmania major* infections. Additionally, the researchers developed and validated a rapid and quantitative biosensor for identifying dengue virus serotypes (1-4) [162]. The biosensor uses cadmium selenide tellurium sulphide (CdSeTeS) fluorescent quantum dots (QDs) and Au NPs in combination with specific hairpin single-stranded DNA probes for each serotype. The biosensor successfully detected target virus DNA in femtomolar concentrations, demonstrating its ability for accurate serotype identification and potential point-of-care diagnostic use. Additionally, Chowdhury et al. developed a new two-way detection method using nanocomposites of Au NPs and nitrogen, sulfur codoped graphene (N, S-G) quantum dots (QDs) for DENV serotype identification and DNA quantification. The authors demonstrated efficient detection of all four DENV serotypes individually within a concentration range of  $10^{-14}$  to  $10^{-6}$  M, with a low LoD (9.4 fM). The sensing platform also depicted satisfactory performance in identifying and quantifying clinically isolated DENV DNA, leading a facile and reliable method for biomedical applications in sensitive and robust sensing probes. Moreover, Shrivastava et al. developed a similar colorimetric sensing platform utilizing Au NPs to detect ampicillin in urine samples with LoD of  $10 \text{ ng mL}^{-1}$  [209]. All measurements were taken using a UV-Visible spectrophotometer of type-1800-Shimadzu-Japan, matching with a 1-cm quartz cell for the identification of ampicillin. In a separate study, Versiani and coauthors developed a sensitive diagnostic test using Au NRs coated with DENV proteins as LSPR sensors [210]. The LSPR platform detected tiny amounts of DENV antibodies and diluted DENV-positive human-sera accurately. The test differentiated the DENV from other flavivirus infections and identified the specific DENV serotype in patients using standard ELISA-plate spectrophotometers. Further, Jeon et al. established a colorimetric-based sensing platform for visual malaria diagnosis using an aptamer and Au NPs [157]. The authors employed specific aptamer pL1

and cationic polymers (PDDA and PAH) to control Au NPs color properties. In the presence of the malaria biomarker (PLDH), Au NPs turn blue due to induced fit binding, allowing detection at low concentrations and high specificity over interfering proteins in just 40 min. The study demonstrated the successful in vitro detection of DENV using a lateral flow assay based on Au NPs. The estimated detection limit achieved was  $5.12 \times 10^2$  PFU, indicating the assay's sensitivity and potential for practical applications in virus detection [211]. Stephen et al. [212] elucidated the potential of electrochemical sensors for the detection of pathogens attributed with NTDs, evaluating the probe selection, nanomaterials, and future applications of these biosensors.

Piezoelectric systems utilize materials capable of inducing an electric-based response when subjected to mechanical oscillation. These platforms present an attractive sensing alternative for NDTs through mass response-type sensors. For instance, Pirich et al. manipulated the use of cost-effective piezoelectric devices, specifically quartz crystal microbalance (QCM) systems, for disease detection [213]. The study involved the sensors with thin films, regarding bacterial cellulose nanocrystals (CN) to enhance the sensitivity as well as specific detection of DENV protein NS1. The immunochip surface, analyzed by AFM, successfully immobilized IgG NS1. Both QCM and QCM-D (with energy dissipation monitoring) demonstrated effective detection of NS1 in serum with only a 10-fold dilution, with respective LoD of  $0.1 \mu\text{g mL}^{-1}$  and  $0.32 \mu\text{g mL}^{-1}$ . These findings proposed that QCM-D and QCM could be potential tool for sensitive, rapid, and low-cost diagnostic assays for DENV fever. Additionally, Ramos et al. developed a sensitive piezoelectric immunosensor for detecting anti-*Leishmania* antibodies in the serum of canine, crucial for diagnosing visceral leishmaniasis (VL) [214]. The sensor used re-combinant antigens immobilized on a quartz crystal electrode coated with a nafion film and Au NPs to improve stability and surface area. The immunosensor depicted an excellent linearity and low relative error at different serum dilutions, outperforming the cysteamine (Cys)-based immunosensor without Au NPs. The study findings demonstrated that the Au NP-based immunosensor held great promise as a reliable tool for VL diagnosis, especially in endemic regions and asymptomatic dog screening, crucial for disease control. In another study, they introduced a novel electrochemical immunosensor based on a one-step fabricated carbon ink graphite screen-printed electrode for detecting non-structural protein NS1 of the reemerging DENV [165]. The sensor's surface is modified using amine-functionalized (-NH<sub>2</sub>) Au NPs synthesized through a photoinduced physical method. The Au NPs improved the sensitivity of NS1 detection by square-wave-voltammetry (SWV) without the need for labeling. The immunosensor demonstrated

a linear response from 0.1 to 2  $\mu\text{g mL}^{-1}$  NS1, with a low LoD (0.03  $\mu\text{g mL}^{-1}$ ). This facile technique showed key potential for early DENV diagnosis, facilitating in controlling its impact on global health. Overall, an in-depth investigation of LSPR based sensors for identification of NTDs can be seen in Table 3.

Responding to the increasing demand for cost-effective, portable, and user-friendly sensing platforms, glass substrates have emerged as a promising avenue for the creation of portable LSPR optical sensors [215]. In this context, Liu and colleagues employed dark-field microscopy to demonstrate that the plasmon shift

of spherical gold nanoparticles immobilized on glass slides can be harnessed to quantitate the presence of 1-propanol, 1-octanol, or oil within a water droplet [216]. More recently, Farooq et al. leveraged Ag NPs on a glass substrate as a foundation for *Candida albicans* sensing [134]. In this study, the Ag NPs of size 10 nm were functionalized with monoclonal IgG anti-*Candida albicans* antibodies for the detection of *Candida albicans* antigen. The practicality of the LSPR sensor was substantiated by successfully identifying *C. albicans* antigen concentrations as low as 50 ng  $\text{mL}^{-1}$  through the use of a spectrophotometer (Ocean Optics HR4000)[134]. In

**Table 3** Selected studies for metallic-based LSPR platforms and their performance

Sensor type	Sample type	Tropical disease	detection limit	References
Optical sensor	Au NPs	Leptospirosis	0.0077 ng/ $\mu\text{L}$	[148]
Optical sensor	Au NPs	Leptospirosis	348 fg/mL	[149]
SERS sensor	Au NPs	Leishmaniasis	-	[150]
Electrochemical sensor	Au NPs	Leishmaniasis	200 ng/mL	[151]
Colorimetric sensor	Au NPs	Leishmaniasis	0.7 pg/ $\mu\text{L}$	[152]
Optical sensor	Ag NPs	Leishmaniasis	4.2 $\mu\text{g/mL}$	[153]
Electrochemical sensor	Ag NPs	Leishmaniasis	0.07 ng/mL	[154]
Electrochemical-based sensor	Ag NPs	Leishmaniasis	0.01 pg/mL	[155]
Electrochemical-based sensor	Ag NPs	Leishmaniasis	29 ng/mL	[156]
Colorimetric-based sensor	Au NPs	Malaria	74 parasites/ $\mu\text{L}$	[157]
PIT-based sensor	Au NPs	Malaria	25 $\mu\text{g/mL}$	[158]
Fluorescence-based sensor	Au NPs	Malaria	1 pg/ $\mu\text{L}$	[159]
Colorimetric-based sensor	Au NPs	Ebola virus	200 ng/mL	[160]
Fluorescence-based sensor	Au NPs	Ebola virus	220 fg/mL	[161]
Fluorescence-based sensor	Au NPs & QDs	DENV (1,2,3,4)	24.6, 11.4, 39.8 and 39.7 fM	[162]
Fluorescence-based sensor	N,S-G QDs & Au NPs	DENV	9.4 pM	[163]
Optical sensor	Au NPs	DENV	0.074 $\mu\text{g/mL}$	[164]
Optical sensor	Au NPs	DENV	0.07 $\mu\text{g/mL}$	[24]
Electrochemical-based sensor	Au NPs	DENV	0.03 $\mu\text{g/mL}$	[165]
Electrochemical-based sensor	Au NPs	DENV	-	[166]
Optical sensor	Au NPs	DENV	2 PFU/mL	[167]
Piezoelectric-based sensor	Au NPs	DENV	0.03 $\mu\text{g/mL}$	[165]
Electrochemical-based sensor	Au NPs	DENV	9.5 $\times$ 10 <sup>5</sup> pfu/mL	[168]
Electrochemical-based sensor	Au NPs	DENV	-	[169]
Optical sensor	Au NPs	DENV	0.07 $\mu\text{g/mL}$	[136]
Optical sensor	Ag NPs	CANDV	50 ng/mL	[134]
Optical sensor	Ag NPs	CANDV	50 ng/mL	[170]
Optical sensor	Ag NPs	CANDV	102 CFU mL	[171]
Geno-sensor	Au NPs	ZIKV	0.2 and 33 fM ng/mL	[172]
Fluorescence-based sensor	Au NPs, QDs	ZIKV	2.9 copies/mL	[173]
Fluorescence sensor	Au NPs, QDs	ZIKV	35.0 copies/mL	[174]
Electrochemical-based sensor	Au NPs	ZIKV	0.82 pmol/L	[175]
SERS-based sensor	Au NPs	ZIKV	10 ng/mL	[176]
Electrochemical-based sensor	Au NPs	ZIKV	10 pM to 1 nM	[177]
Geno-sensor	Au NPs	Schistosomiasis	0.685 pg/mL	[178]
Geno-sensor	Au NPs	Schistosomiasis	0.6 pg/mL	[179]

another study, the enzymatic-like activity of Au NPs, akin to that of enzymes, is intricately influenced by factors such as their aggregation state, surface ligands, and morphology. By harnessing chromogenic substrates, colorimetric sensing platform that capitalizes on the peroxidase-mimicking capacity of Au NPs can be proficiently employed for the purpose of pathogen detection [217]. Additionally, a new diagnostic method was developed by Camara et al. for dengue fever in the acute phase of infection [164]. The method applies on an all-optical fiber sensor based on LSPR and specular reflection from Au NPs to detect NS1 antigen at different concentrations. The optical fiber-based sensor holds promise as a potential tool for DENV diagnosis with LoD of 0.074  $\mu\text{g/mL}$  due to its rapid, reliable, and user-friendly simple operation. In an alternative approach, the researchers introduced novel label-free LSPR immunosensors based on Au NPs for DENV sensing. By optimizing spherical-shaped Au NPs size, the platforms show high molecular sensitivity, a sharp spectral resonance peak, and a high figure of merit (FoM). Further, the their platforms demonstrate successful identification of dengue NS1 antigens with a low limit of detection, presenting promise for engineering efficient immunosensor platforms [24, 136]. Notably, the advantages and disadvantages of LSPR based sensors can be found in Table 4.

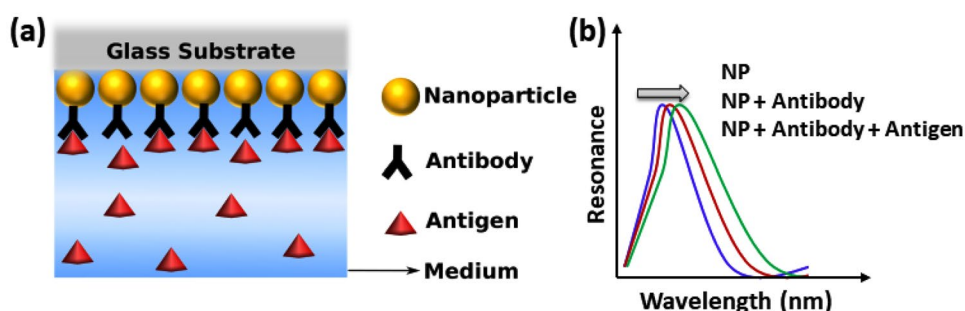
An alternative strategy for enhancing both robust LSPR shift and sensing specificity involves employing a “sandwich assay” with an additional antigen or antibody molecule. Recently, Deroco et al. [73] showcased the use of a sandwich-immunoassay LSPR sensing platform to diagnose NTDs. A new assay using lambda exonuclease ( $\lambda$

-exonuclease) and SERS was introduced for fast detection of three bacterial meningitis pathogens in cerebral spinal fluid. The assay showed high sensitivity, multiplexing capability, and quantification of individual pathogens using unique SERS signals merged with partial least squares (PLS) regression. This innovative SERS assay offers significant advantages over current fluorescent-based methods and presents promising opportunities for future advancement in healthcare management, ensuring timely and effective diagnosis of bacterial meningitis [218]. A colorimetric sensing method using dual amplification was employed for detecting thrombin. The assay involved two aptamers (TBA1 and TBA2) in a sandwich configuration with Au NPs [219]. The transition from a red to purple hue upon thrombin binding to TBA1, coupled with the redshift observed in the absorption peak due to the integration of TBA2 with Au NPs, assisted the naked-eye identification of thrombin concentrations at an ultralow level of 3  $\mu\text{g mL}^{-1}$ . Employing UV-Vis analysis, LoD was rigorously determined to be 1.33  $\mu\text{g mL}^{-1}$ . Impressively, the sensing platform displayed exceptional discernment even in the presence of intricate interferents, yielding a remarkable recovery rate of 97.4% when assessed with authentic biological human serum samples. The application, furthermore, of the sandwich-immunoassay LSPR technique was extended to the detection of the hepatitis-B virus surface antigen (HBsAg). In a novel study, Kim et al. established a glass substrate featuring Au NPs coupled with an anti-HBsAg antibody, enabling for the detection of HBsAg through the development of a sandwich immunoassay involving an additional layer of Au NPs functionalized with anti-HBsAg antibodies. This innovative approach,

**Table 4** Advantages and disadvantages of sensors, biosensors, and conventional methods with limitations

Aspect	Sensors	Biosensors	Conventional methods
<b>Advantages</b>	<ul style="list-style-type: none"> <li>- High sensitivity</li> <li>- Real-time monitoring</li> <li>- Low sample volume</li> <li>- Minimal sample handling</li> <li>- Potential for automation</li> </ul>	<ul style="list-style-type: none"> <li>- High specificity</li> <li>- Rapid detection</li> <li>- Quantitative data</li> <li>- Minimal sample preparation</li> <li>- Potential for miniaturization</li> <li>- Integrated with electronics</li> </ul>	<ul style="list-style-type: none"> <li>- Established methods</li> <li>- Widely availability</li> <li>- Established expertise</li> <li>- Well-recognized</li> <li>- Cost-effective</li> </ul>
<b>Disadvantages</b>	<ul style="list-style-type: none"> <li>- Specific to target analytes</li> <li>- Limited specificity</li> <li>- Cost of instrumentation</li> <li>- Expertise required</li> <li>- Limited multiplexing</li> </ul>	<ul style="list-style-type: none"> <li>- Design complexity</li> <li>- Cost of bio-receptors</li> <li>- Stability of bioreceptors</li> <li>- Potential for false positives</li> <li>- Limited multiplexing</li> </ul>	<ul style="list-style-type: none"> <li>- Limited sensitivity</li> <li>- Limited specificity</li> <li>- Labor-intensive</li> <li>- Long turnaround time</li> <li>- Expensive equipment</li> </ul>
<b>Limitations</b>	<ul style="list-style-type: none"> <li>- Interference from matrix</li> <li>- Calibration requirements</li> <li>- Biological fouling</li> <li>- Complex signal analysis</li> </ul>	<ul style="list-style-type: none"> <li>- Stability of recognition</li> <li>- Potential cross-reactivity</li> <li>- Shelf life of bioreceptors</li> <li>- Sensitivity to environmental conditions</li> <li>- Limited target range</li> </ul>	<ul style="list-style-type: none"> <li>- Sensitivity challenges</li> <li>- Lack of specificity</li> <li>- Sample variability</li> <li>- Infrastructure challenges</li> </ul>

**Fig. 9** **a** The schematic diagram of the LSPR sensing platform used for molecular detection. **b** The plasmonic spectral peak of NPs as a function of molecular interaction with NPs, i.e., antibody and antigen [123].



as detailed by Kim et al. [220], effectively enhanced the platform's LoD.

In another exceptional techniques for sensing, Nasrin et al. introduced a new dual-mode detection platform for recombinant of CHIKV E1 protein using NPs. The system integrated both fluorescence and electrochemical signals for detection [221]. This method included immobilizing NPs with methylene blue (MB) inside liposomes, and after adding CHIKV, the sandwich structure was magnetically isolated using antibodies and modified  $Fe_3O_4$  NPs. By releasing the nanoconjugates and monitoring the fluorescence and electrochemical signals, CHIKV was detected at a low concentration, with a LoD of  $32 \text{ fg mL}^{-1}$ . Such devices explore changes in plasmonic resonance peaks due to the adsorption of specific molecules or biomarkers (Fig. 9). Importantly, this biosensor obtains a selective LoD of  $100 \text{ pg mL}^{-1}$ , showing exceptional potential for affordable, portable diagnostics across critical diseases.

## Challenges and Limitations

### Stability

The stability of plasmonic nanoparticles (NPs) is crucial for their effective utilization in various sensing applications, as their high surface-to-volume ratio makes them prone to aggregation, leading to a loss of their unique properties. Researchers employ surface modification techniques, such as the use of stabilizing agents, to prevent aggregation and enhance the potential of these NPs in fields like nanomedicine and catalysis. In the realm of molecular diagnostics, plasmonic NPs have been extensively studied for LSPR colorimetric assays [207]; however, the risk of NP aggregation due to non-target factors like ionic strength, pH, solvent composition, and temperature can result in false-negative or false-positive outcomes. Strategies to mitigate aggregation include exploiting the selectivity of aggregation induced by specific factors, such as mercury ions. Additionally, techniques like laser ablation are employed to avoid aggregation during NP synthesis, providing NPs with a negative

zeta potential that enhances their stability even at room temperatures. For instance, TiN NPs synthesized through laser ablation in colloids have shown promise for long-term colloidal stability due to their negative charge, contributing to extended electrostatic stabilization of the solutions [222].

### Reproducibility

The reproducibility of NPs poses a huge challenge in optical sensing applications. The synthesis of NPs in suspension includes complex chemical reactions in thermodynamically and kinetically controlled conditions, and small variations in reaction conditions can lead to differences in shape, size, as well as compositions of the NPs, exploiting their characteristics and behavior. Consequently, NPs' high surface area can result in agglomeration and sedimentation, further impacting their reproducibility [223]. Diverse characterization techniques, such as SEM, TEM, and DLS, could yield slightly different outcomes, contributing to variability in reported properties [224]. On large-scale production, providing consistent quality and reproducibility across huge batches is challenging due to manufacturing processes, fluctuations in raw materials, or equipment. Additionally, the surface chemistry and properties of NPs could be sensitive to environmental conditions and interactions with other molecules, enabling to variations in their performance. Over time, NPs may also exhibit changes in size, shape, or aggregation, adding to the complexity of achieving reproducible results. Fortunately, achieving this goal is entirely feasible and within reach. Addressing these challenges demands standardized protocols, careful experimental design, and thorough characterization to ensure reliable and reproducible NP synthesis. Evaluating these several interfaces ensures the advancement of predictive relations between structure and activity, determined by nanomaterial features like shape, size, roughness, surface chemistry, as well as surface coatings. This type of knowledge is fruitful regarding safe use of nanomaterials. Further, integrated efforts and open data sharing are the key components among researchers, can support validation, and establish strong guidelines for NP production and characterization, leading to broad reproducibility in this dynamic field [225].

## Nanomaterials' Effects on Ecosystem

The broad range of use of NPs across multiple applications has alarmed about their adverse effects on the environment, and living organisms, including humans. A plethora of research has shown that prolonged exposure to these NPs may exhibit to both short and long-term toxicity [226–228]. Even in non-toxic form, Ag NPs can be harmful to humans, damaging cell membranes, mitochondria, and DNA, resulting in structural malfunctions and apoptosis [229, 230]. Furthermore, inhalation of NPs can lead to their deposition in the nasopharyngeal vein, providing a pathway for them to reach the cerebellum via olfactory nerves. Because of their small size, these NPs can penetrate the blood-brain barrier and result in neurotoxicity and dopamine depletion in brain cells, potentially affecting brain physiology and function [231]. These findings show the importance of understanding and addressing the potential risks attributed to the use of NPs in various applications.

## Quantification

While the LSPR sensor design has advanced significantly, the practical application of SERS detection in areas requiring high accuracy, like sensing and medical diagnosis, remains limited due to the huge challenge of quantification [232]. The SERS offers remarkable signal enhancement, enabling single-molecule-level detection, it faces limitations related to specificity and reliability due to the variability of enhancement factors ( $EF_{SERS}$ ) for different analytes and NPs. Particularly at ultra-low analyte concentrations ( $< 10^{-7} molL^{-1}$ ), the probability of achieving significant SERS enhancement is low, leading to temporal fluctuations in intensity and spectral position, known as 'blinking', which poses challenges for molecular recognition [233]. Researchers have proposed mechanisms such as adsorption, molecular diffusion cycles, and changes in hotspot configuration as potential causes of these instabilities.

## Future Directions

The future of LSPR biosensors for NTDs promises to revolutionize disease detection, diagnosis, and healthcare management. Researchers are focused on improving sensitivity and selectivity, aiming to identify specific biomarkers associated with tropical diseases accurately. Through nanostructure engineering and functionalization optimization, these advancements will enable precise and reliable disease diagnosis, even at low analyte concentrations. Additionally, the development of multiplexing capabilities allows for the simultaneous detection of multiple disease indicators in a single sample, expediting diagnosis and identifying co-infections prevalent in tropical regions.

The future of LSPR biosensors also entails the creation of field-deployable and robust designs, essential for regions with limited access to modern laboratory facilities. Portable, user-friendly point-of-care diagnostic devices will bring diagnostics closer to patients, particularly in resource-limited tropical areas, enabling quicker intervention and alleviating the burden on healthcare infrastructure. Furthermore, the affordability and accessibility of LSPR biosensors will facilitate large-scale disease monitoring and surveillance in tropical settings, empowering public health officials to implement targeted interventions effectively.

Integration with emerging technologies, such as microfluidics, machine learning algorithms, and smartphone connectivity, is anticipated to enhance biosensor performance, streamline data processing, and enable remote monitoring capabilities. Collaborative research efforts and international partnerships are essential for accelerating progress in this field, enabling transformative solutions to combat tropical diseases and improve global health outcomes. Overall, the future of LSPR biosensors for tropical illnesses aims to enhance accessibility, efficiency, and impact in the fight against infectious diseases prevalent in tropical regions, ultimately benefiting vulnerable populations.

## Conclusion

In conclusion, this study underscores the pivotal role of metallic-based LSPR sensors in tropical disease detection, particularly in the context of Neglected Tropical Diseases (NTDs). These sensors offer high sensitivity and label-free detection, making them valuable tools for swift and accurate diagnosis. The integration of LSPR sensors with smartphone technology enhances point-of-care testing, providing cost-effective solutions that are particularly beneficial in regions with limited healthcare infrastructure.

Recent research has demonstrated the potential of metallic-based LSPR sensors in detecting a range of tropical illnesses, including Chagas disease, leishmaniasis, dengue fever, Zika virus infection, and sleeping sickness. These sensors have the capacity to significantly improve disease diagnosis and treatment. Furthermore, their adaptability to complex biological environments and the ability to provide rapid results make them promising contributors to global health initiatives aimed at combating NTDs.

Overall, metallic-based LSPR sensors demonstrate a transformative technology in the diagnosis and monitoring of tropical diseases. Their exceptional attributes, such as high sensitivity and portability, make them invaluable tools for enhancing healthcare accessibility and improving overall healthcare outcomes in regions burdened by tropical illnesses. The integration of LSPR sensors into healthcare

systems holds the potential to drive earlier detection and more effective treatment, making a substantial impact on global efforts to combat NTDs.

**Funding** This work was funded by CNPq (INCT-INTERAS 406761/2022-1), INCT-INFO (465763/2014-6); Sisfoton (440228/2021-2); PQ (314517/2021-9); CAPES Finance code 001 and FAPESP (17/50332-0) and FAPESP (21/00633-0).

**Data Availability** The data is available on request from the corresponding author.

## Declarations

**Conflict of Interest** The authors declare no competing interests.

## References

- Lee S, Song H, Ahn H, Kim S, Jr Choi, Kim K (2021) Fiber-optic localized surface plasmon resonance sensors based on nanomaterials. *Sensors* 21(3):819
- Xu G, Du X, Wang W, Qu Y, Liu X, Zhao M, Li W, Li YQ (2022) Plasmonic nanozymes: leveraging localized surface plasmon resonance to boost the enzyme-mimicking activity of nanomaterials. *Small* 18(49):2204131
- Min Y, Wang Y (2020) Manipulating bimetallic nanostructures with tunable localized surface plasmon resonance and their applications for sensing. *Front Chem* 8:411
- Hong Y, Reinhard BM (2019) Optoplasmonics: basic principles and applications. *J Opt* 21(11):113001
- Farooq S, Shafique S, Ahsan Z, Cardozo O, Wali F (2022a) Tailoring the scattering response of optical nanocircuits using modular assembly. *Nanomaterials* 12(17):2962
- Khurana K, Jaggi N (2021) Localized surface plasmonic properties of Au and Ag nanoparticles for sensors: a review. *Plasmonics* 16(4):981–999
- Yadav A, Gerislioglu B, Ahmadvand A, Kaushik A, Cheng GJ, Ouyang Z, Wang Q, Yadav VS, Mishra YK, Wu Y et al (2021) Controlled self-assembly of plasmon-based photonic nanocrystals for high performance photonic technologies. *Nano Today* 37:101072
- Janith G, Herath H, Hendeniya N, Attygalle D, Amarasinghe D, Logeeshan V, Wickramasinghe P, Wijayasinghe Y (2023) Advances in surface plasmon resonance biosensors for medical diagnostics: an overview of recent developments and techniques. *Journal of Pharmaceutical and Biomedical Analysis Open* 2:100019
- Siddique S, Chow JC (2020) Application of nanomaterials in biomedical imaging and cancer therapy. *Nanomaterials* 10(9):1700
- Balitskii O (2021) Recent energy targeted applications of localized surface plasmon resonance semiconductor nanocrystals: a mini-review. *Mater Today Energy* 20:100629
- Kim DM, Park JS, Jung SW, Yeom J, Yoo SM (2021a) Biosensing applications using nanostructure-based localized surface plasmon resonance sensors. *Sensors* 21(9):3191
- Mie G (1908) Optical characteristics of turbid tubes, especially colloidal metal solutions. *Ann Phys* 330(3):377–445. <https://doi.org/10.1002/andp.19083300302>
- Prodan E, Radloff C, Halas NJ, Nordlander P (2003) A hybridization model for the plasmon response of complex nanostructures. *Science* 302(5644):419–422
- Sarkar S, König TA (2023) Engineering plasmonic hybridization toward advanced optical sensors. *Advanced Sensor Research* 2300054
- Jee Y, Yu Y, Abernathy HW, Lee S, Kalapos TL, Hackett GA, Ohodnicki PR (2018) Plasmonic conducting metal oxide-based optical fiber sensors for chemical and intermediate temperature-sensing applications. *ACS Appl Mater Interfaces* 10(49):42552–42563. <https://doi.org/10.1021/acsami.8b11956>
- Jadhav P, Muhammad N, Bhuyar P, Krishnan S, Abd Razak AS, Zularisam A, Nasrullah M (2021) A review on the impact of conductive nanoparticles (cnps) in anaerobic digestion: applications and limitations. *Environ Technol Innov* 23:101526
- Garcia-Vidal FJ, Fernández-Domínguez AI, Martín-Moreno L, Zhang HC, Tang W, Peng R, Cui TJ (2022) Spoof surface plasmon photonics. *Rev Mod Phys* 94(2):025004
- Mittal S, Sharma T, Tiwari M (2021) Surface plasmon resonance based photonic crystal fiber biosensors: a review. *Mater Today Proc* 43:3071–3074
- Hutter E, Fendler JH (2004) Exploitation of localized surface plasmon resonance. *Adv Mater* 16(19):1685–1706
- Farooq S, de Araujo RE (2018) Engineering a localized surface plasmon resonance platform for molecular biosensing. *Open J Appl Sci* 8(3):126–139
- Badilescu S, Raju D, Bathini S, Packirisamy M (2020) Gold nano-island platforms for localized surface plasmon resonance sensing: a short review. *Molecules* 25(20):4661
- Todorov R, Hristova-Vasileva T, Katrova V, Atanasova A (2023) Silver and gold containing compounds of p-block elements as perspective materials for UV plasmonics. *ACS Omega* 8(16):14321–14341
- Gurav DD, Jia YA, Ye J, Qian K (2019) Design of plasmonic nanomaterials for diagnostic spectrometry. *Nanoscale Adv* 1(2):459–469. <https://doi.org/10.1039/C8NA00319J>
- Farooq S, Wali F, Zzell DM, de Araujo RE, Rativa D (2022b) Optimizing and quantifying gold nanospheres based on Ispr label-free biosensor for dengue diagnosis. *Polymers* 14(8):1592
- Lee SA, Link S (2021) Chemical interface damping of surface plasmon resonances. *Acc Chem Res* 54(8):1950–1960
- Jeon HB, Tsalu PV, Ha JW (2019) Shape effect on the refractive index sensitivity at localized surface plasmon resonance inflection points of single gold nanocubes with vertices. *Sci Rep* 9(1):13635
- Zhang J, Kolhatkar G, Ruediger A (2021) Localized surface plasmon resonance shift and its application in scanning near-field optical microscopy. *J Mater Chem C* 9(22):6960–6969
- Xu T, Geng Z (2021) Strategies to improve performances of Ispr biosensing: structure, materials, and interface modification. *Biosens Bioelectron* 174:112850
- Hong YA, Ha JW (2022) Enhanced refractive index sensitivity of localized surface plasmon resonance inflection points in single hollow gold nanospheres with inner cavity. *Sci Rep* 12(1):6983
- Martín-Sánchez C, Sánchez-Iglesias A, Barrera-Argüeso JA, Polian A, Liz-Marzán LM, Rodríguez F (2022) Behavior of Au nanoparticles under pressure observed by in situ small-angle x-ray scattering. *ACS Nano* 17(1):743–751
- Chen Y, Ai B, Wong ZJ (2020) Soft optical metamaterials. *Nano. Convergence* 7:1–17
- Katyal J et al (2021) Localized surface plasmon resonance and field enhancement of Au, Ag, Al and Cu nanoparticles having isotropic and anisotropic nanostructure. *Mater Today Proc* 44:5012–5017
- Guvenc CM, Balci FM, Sarisozen S, Polat N, Balci S (2020) Colloidal bimetallic nanorings for strong plasmon exciton coupling. *J Phys Chem C* 124(15):8334–8340
- Balci FM, Sarisozen S, Polat N, Guvenc CM, Karadeniz U, Tertemiz A, Balci S (2021) Laser assisted synthesis of anisotropic metal nanocrystals and strong light-matter coupling in decahedral bimetallic nanocrystals. *Nanoscale Advances* 3(6):1674–1681

35. Do PQT, Huong VT, Phuong NTT, Nguyen TH, Ta HKT, Ju H, Phan TB, Phung VD, Tran NHT et al (2020) The highly sensitive determination of serotonin by using gold nanoparticles (aunps) with a localized surface plasmon resonance (lspr) absorption wavelength in the visible region. *RSC Adv* 10(51):30858–30869
36. Dahlman CJ, Agrawal A, Staller CM, Adair J, Milliron DJ (2019) Anisotropic origins of localized surface plasmon resonance in n-type anatase tio<sub>2</sub> nanocrystals. *Chem Mater* 31(2):502–511
37. Yao M, Ning D, Lin X, Huang J, Huang S, Lin T, Zou B, Hong P, Liang Y (2023) Tunable surface plasmonic resonance and infrared self-focusing propagation in cuxs nanoparticle suspensions. *Opt Commun* 527
38. Tanjaya NK, Kaur M, Nagao T, Ishii S (2022) Photothermal heating and heat transfer analysis of anodic aluminum oxide with high optical absorptance. *Nanophotonics* 11(14):3375–3381
39. Cardozo O, Farooq S, Farias PM, Fraidenaich N, Stingl A, Araujo RE (2022) Zinc oxide nanodiffusers to enhance p3ht:pcbm organic solar cells performance. *J Mater Sci Mater Electron* 33(6):3225–3236
40. Farooq S, Rativa D, de Araujo RE (2021) High performance gold dimeric nanorods for plasmonic molecular sensing. *IEEE Sens J* 21(12):13184–13191
41. Piotta V, Litti L, Meneghetti M (2020) Synthesis and shape manipulation of anisotropic gold nanoparticles by laser ablation in solution. *J Phys Chem C* 124(8):4820–4826
42. Bu Y, Huang R, Li Z, Zhang P, Zhang L, Yang Y, Liu Z, Guo K, Gao F (2021) Anisotropic truncated octahedral au with pt deposition on arris for localized surface plasmon resonance-enhanced photothermal and photodynamic therapy of osteosarcoma. *ACS Appl Mater Interfaces* 13(30):35328–35341
43. González-Rubio G, Scarabelli L, Guerrero-Martínez A, Liz-Marzán LM (2020) Surfactant-assisted symmetry breaking in colloidal gold nanocrystal growth. *ChemNanoMat* 6(5):698–707
44. Farooq S, Nunes FD, de Araujo RE (2018c) Optical properties of silver nanoplates and perspectives for biomedical applications. *Photonics Nanostruct Fundam Appl* 31:160–167
45. Martínez Castellano E, Tamayo-Arriola J, Montes Bajo M, Gonzalo A, Stanojević L, Ulloa JM, Klymov O, Yeste J, Agouram S, Muñoz E et al (2021) Self-assembled metal-oxide nanoparticles on gaas: infrared absorption enabled by localized surface plasmons. *Nanophotonics* 10(9):2509–2518
46. Kasani S, Zheng P, Bright J, Wu N (2019) Tunable visible-light surface plasmon resonance of molybdenum oxide thin films fabricated by e-beam evaporation. *ACS Appl Electron Mater* 1(11):2389–2395
47. Duque JS, Madrigal BM, Riascos H, Avila YP (2019) Colloidal metal oxide nanoparticles prepared by laser ablation technique and their antibacterial test. *Colloids Interfaces* 3(1):25
48. Proenca M, Borges J, Rodrigues MS, Meira DI, Sampaio P, Dias JP, Pedrosa P, Martin N, Bundaleski N, Teodoro OM et al (2019) Nanocomposite thin films based on au-ag nanoparticles embedded in a cuo matrix for localized surface plasmon resonance sensing. *Appl Surf Sci* 484:152–168
49. Chowdhury AD, Takemura K, Khorish IM, Nasrin F, Tun MMN, Morita K, Park EY (2020a) The detection and identification of dengue virus serotypes with quantum dot and aunp regulated localized surface plasmon resonance. *Nanoscale Advances* 2(2):699–709
50. Son J, Choi D, Park M, Kim J, Jeong KS (2020) Transformation of colloidal quantum dot: from intraband transition to localized surface plasmon resonance. *Nano Lett* 20(7):4985–4992
51. Mousavi SM, Hashemi SA, Kalashgrani MY, Rahmanian V, Gholami A, Chiang WH, Lai CW (2022) Biomedical applications of an ultra-sensitive surface plasmon resonance biosensor based on smart mxene quantum dots (smqds). *Biosensors* 12(9):743
52. Li H, Xu Q, Wang X, Liu W (2018) Ultrasensitive surface-enhanced raman spectroscopy detection based on amorphous molybdenum oxide quantum dots. *Small* 14(28):1801523
53. Yakimchuk DV, Kaniukov EY, Lepeshov S, Bundyukova VD, Demyanov SE, Arzumanyan GM, Doroshkevich NV, Mamatkulov KZ, Bochmann A, Presselt M, et al. (2019) Self-organized spatially separated silver 3d dendrites as efficient plasmonic nanostructures for surface-enhanced raman spectroscopy applications. *J Appl Phys* 126(23)
54. Giordano MC, Tzschoppe M, Barelli M, Vogt J, Huck C, Canepa F, Pucci A, Buatier de Mongeot F (2020) Self-organized nanorod arrays for large-area surface-enhanced infrared absorption. *ACS Appl Mater Interfaces* 12(9):11155–11162
55. Germanicus RC, Bourlier Y, Notot V, Bérimi B, Demange V, Berthe M, Boileau A, Euchin M, Dumont Y, Aureau D et al (2020) Three dimensional resistance mapping of self-organized sr<sub>3</sub>v<sub>2</sub>o<sub>8</sub> nanorods on metallic perovskite srvo<sub>3</sub> matrix. *Appl Surf Sci* 510:145522
56. Tim B, Błaszkievicz P, Kotkowiak M (2021) Recent advances in metallic nanoparticle assemblies for surface-enhanced spectroscopy. *Int J Mol Sci* 23(1):291
57. Alharbi R, Irannejad M, Yavuz M (2019) A short review on the role of the metal-graphene hybrid nanostructure in promoting the localized surface plasmon resonance sensor performance. *Sensors* 19(4):862
58. Liu C, Chen H, Wang S, Liu Q, Jiang YG, Zhang DW, Liu M, Zhou P (2020) Two-dimensional materials for next-generation computing technologies. *Nat Nanotechnol* 15(7):545–557
59. Ghopry SA, Alamri MA, Goul R, Sakidja R, Wu JZ (2019) Extraordinary sensitivity of surface-enhanced Raman spectroscopy of molecules on MoS<sub>2</sub> (WS<sub>2</sub>) nanodomes/graphene Van der Waals heterostructure substrates. *Adv Opt Mater* 7(8):1801249. <https://doi.org/10.1002/adom.201801249>
60. Rohaizad N, Mayorga-Martinez CC, Fojt M, Latiff NM, Pumera M (2021) Two-dimensional materials in biomedical, biosensing and sensing applications. *Chem Soc Rev* 50(1):619–657
61. Fernández-Arias M, Boutinguiza M, del Val J, Riveiro A, Rodríguez D, Arias-González F, Gil J, Pou J (2020) Fabrication and deposition of copper and copper oxide nanoparticles by laser ablation in open air. *Nanomaterials* 10(2):300
62. Machado TM, Peixoto LPF, Andrade GF, Silva MA (2022) Copper nanoparticles-containing tellurite glasses: an efficient sers substrate. *Mater Chem Phys* 278:125597
63. Popok VN, Novikov SM, Lebedinskij YY, Markeev AM, Andreev AA, Trunkin IN, Arsenin AV, Volkov VS (2021) Gas-aggregated copper nanoparticles with long-term plasmon resonance stability. *Plasmonics* 16:333–340
64. Dong C, Feng W, Xu W, Yu L, Xiang H, Chen Y, Zhou J (2020) The coppery age: copper (cu)-involved nanotheranostics. *Adv Sci* 7(21):2001549
65. Ballesteros CA, Correa DS, Zucolotto V (2020) Polycaprolactone nanofiber mats decorated with photoresponsive nanogels and silver nanoparticles: slow release for antibacterial control. *Mater Sci Eng C* 107
66. Amirjani A, Firouzi F, Haghshenas DF (2020) Predicting the size of silver nanoparticles from their optical properties. *Plasmonics* 15:1077–1082
67. Krishnan PD, Banas D, Durai RD, Kabanov D, Hosnedlova B, Kepinska M, Fernandez C, Ruttkay-Nedecky B, Nguyen HV, Farid A et al (2020) Silver nanomaterials for wound dressing applications. *Pharmaceutics* 12(9):821
68. Lee JH, Cho HY, Choi HK, Lee JY, Choi JW (2018) Application of gold nanoparticle to plasmonic biosensors. *Int J Mol Sci* 19(7):2021
69. He MQ, Yu YL, Wang JH (2020) Biomolecule-tailored assembly and morphology of gold nanoparticles for lspr applications. *Nano Today* 35:101005

70. Pedrosa TdL, Farooq S, de Araujo RE (2022) Selecting high-performance gold nanorods for photothermal conversion. *Nanomaterials* 12(23):4188
71. Wang Z, Ren X, Wang D, Guan L, Li X, Zhao Y, Liu A, He L, Wang T, Zvyagin AV et al (2023) Novel strategies for tumor radiosensitization mediated by multifunctional gold-based nanomaterials. *Biomater Sci* 11:1116–1136
72. Souto DE, Volpe J, Gonçalves CdC, Ramos CH, Kubota LT (2019a) A brief review on the strategy of developing spr-based biosensors for application to the diagnosis of neglected tropical diseases. *Talanta* 205:120122
73. Deroco PB, Junior DW, Kubota LT (2021) Recent advances in point-of-care biosensors for the diagnosis of neglected tropical diseases. *Sens Actuators, B Chem* 349:130821
74. Xifre-Perez E, Ferre-Borrull J, Marsal LF (2022) Oligonucleotide probes and immunosensors based on nanoporous anodic alumina for screening of diseases. *Adv Mater Technol* 7(9):2101591
75. Souto DE, Volpe J, Gonçalves CdC, Ramos CH, Kubota LT (2019b) A brief review on the strategy of developing spr-based biosensors for application to the diagnosis of neglected tropical diseases. *Talanta* 205:120122
76. de Freitas Borges PA, Fiel WA, Vasconcellos VA, de Faria RAD (2021) Current progresses in the development of biosensors for the diagnosis of neglected tropical diseases. *Systematic Bioscience and Engineering* pp 39–49
77. Samuel VR, Rao KJ (2022) A review on label free biosensors. *Biosens Bioelectron*: X p 100216
78. Zheng Y, Bian S, Sun J, Wen L, Rong G, Sawan M (2022) Label-free lspr-vertical microcavity biosensor for on-site sars-cov-2 detection. *Biosensors* 12(3):151
79. Taghavi A, Rahbarizadeh F, Abbasian S, Moshaii A (2020) Label-free lspr prostate-specific antigen immune-sensor based on glads-fabricated silver nano-columns. *Plasmonics* 15:753–760
80. Erdem Ö, Saylan Y, Cihangir N, Denizli A (2019) Molecularly imprinted nanoparticles based plasmonic sensors for real-time enterococcus faecalis detection. *Biosens Bioelectron* 126:608–614
81. Zhu S, Xie Z, Chen Y, Liu S, Kwan YW, Zeng S, Yuan W, Ho HP (2022) Real-time detection of circulating tumor cells in blood-stream using plasmonic fiber sensors. *Biosensors* 12(11):968
82. Kim HM, Park JH, Jeong DH, Lee HY, Lee SK (2018a) Real-time detection of prostate-specific antigens using a highly reliable fiber-optic localized surface plasmon resonance sensor combined with micro fluidic channel. *Sens Actuators B* 273:891–898
83. Roether J, Chu KY, Willenbacher N, Shen AQ, Bhalla N (2019) Real-time monitoring of dna immobilization and detection of dna polymerase activity by a microfluidic nanoplasmonic platform. *Biosens Bioelectron* 142:111528
84. Chen H, Liu K, Li Z, Wang P (2019) Point of care testing for infectious diseases. *Clin Chim Acta* 493:138–147
85. Hoque S, Somasundaram L, Samy R, Dawane A, Sen A (2021) Localized surface plasmon resonance sensors for biomarker detection with on-chip microfluidic devices in point-of-care diagnostics. In: *Advanced Micro-and Nano-Manufacturing Technologies: Applications in Biochemical and Biomedical Engineering*, Springer, pp 199–223
86. Zhu L, Ling J, Zhu Z, Tian T, Song Y, Yang C (2021) Selection and applications of functional nucleic acids for infectious disease detection and prevention. *Anal Bioanal Chem* 413:4563–4579
87. Jampasa S, Kreangkaiwal C, Kalcher K, Waiwinya W, Techawattananaboon T, Songumpai N, Sueyanyongsiri P, Pattanasombatsakul K, Techapornroong M, Benjamanukul S et al (2022) Resistance-based lateral flow immunosensor with a nfc-enabled smartphone for rapid diagnosis of leptospirosis in clinical samples. *Anal Chem* 94(42):14583–14592
88. Figueredo F, Stolowicz F, Vojnov A, Coltro WK, Larocca L, Carrillo C, Cortón E (2021) Towards a versatile and economic Chagas disease point-of-care testing system, by integrating loop-mediated isothermal amplification and contactless/label-free conductivity detection. *PLoS Negl Trop Dis* 15(5)
89. Prado IC, Mendes VG, Souza AL, Dutra RF, De-Simone SG (2018) Electrochemical immunosensor for differential diagnostic of wuchereria bancrofti using a synthetic peptide. *Biosens Bioelectron* 113:9–15
90. Awan M, Rauf S, Abbas A, Nawaz MH, Yang C, Shahid SA, Amin N, Hayat A (2020) A sandwich electrochemical immunosensor based on antibody functionalized-silver nanoparticles (ab-ag nps) for the detection of dengue biomarker protein ns1. *J Mol Liq* 317:114014
91. Lee JS, Kim J, Shin H, Min DH (2020) Graphene oxide-based molecular diagnostic biosensor for simultaneous detection of zika and dengue viruses. *2D Materials* 7(4):044001
92. Booth M (2018) Climate change and the neglected tropical diseases. *Adv Parasitol* 100:39–126
93. Gutman JR, Lucchi NW, Cantey PT, Steinhardt LC, Samuels AM, Kamb ML, Kapella BK, McElroy PD, Udhayakumar V, Lindblade KA (2020) Malaria and parasitic neglected tropical diseases: potential syndemics with COVID-19? *Am J Trop Med Hyg* 103(2):572
94. Molyneux DH, Aboe A, Isiyaku S, Bush S (2020) COVID-19 and neglected tropical diseases in Africa: impacts, interactions, consequences. *Int Health* 12(5):367–372
95. da Conceição JR, Lopes CPG, Ferreira EI, Epiphanyo S, Giarolla J (2022) Neglected tropical diseases and systemic racism especially in Brazil: from socio-economic aspects to the development of new drugs. *Acta Trop* 235
96. Bryson JM, Bishop-Williams KE, Berrang-Ford L, Nunez EC, Lwasa S, Namanya DB, Harper SL, Team IHATCCR, et al. (2020) Neglected tropical diseases in the context of climate change in east Africa: a systematic scoping review. *Am J Trop Med Hyg* 102(6):1443
97. Morgan J, Strode C, Salcedo-Sora JE (2021) Climatic and socio-economic factors supporting the co-circulation of dengue, zika and chikungunya in three different ecosystems in colombia. *PLoS Negl Trop Dis* 15(3):e0009259
98. Telle O, Nikolay B, Kumar V, Benkimoun S, Pal R, Nagpal B, Paul RE (2021) Social and environmental risk factors for dengue in Delhi city: a retrospective study. *PLoS Negl Trop Dis* 15(2)
99. Lima D, Hacke ACM, Ulmer B, Kuss S (2021) Electrochemical sensing of trypanosome-and flavivirus-related neglected tropical diseases. *Curr Opin Electrochem* 30:100838
100. Yang H, Ledesma-Amaro R, Gao H, Ren Y, Deng R (2023a) Crispr-based biosensors for pathogenic biosafety. *Biosensors and Bioelectronics* p 115189
101. Zakiyyah SN, Ibrahim AU, Babiker MS, Gaffar S, Ozsoz M, Zein MIH, Hartati YW (2022) Detection of tropical diseases caused by mosquitoes using crispr-based biosensors. *Tropical Medicine and Infectious Disease* 7(10):309
102. Castro-Camus E, Koch M, Mittleman DM (2022) Towards the development of thz-sensors for the detection of African trypanosomes. *Appl Phys B* 128(1):12
103. Knieß R, Wagner CB, Ulrich Göringer H, Mueh M, Damm C, Sawallich S, Chmielak B, Plachetka U, Lemme M (2018) Towards the development of thz-sensors for the detection of African trypanosomes. *Frequenz* 72(3–4):101–111
104. Kim DM, Park JS, Jung SW, Yeom J, Yoo SM (2021b) Biosensing applications using nanostructure-based localized surface plasmon resonance sensors. *Sensors* 21(9):3191
105. Lim SG, Jo S, Lee JH, Kwona OS (2022) Review for device compositions of localized surface plasmon resonance sensors. *Appl Sci Converg Technol* 31(2):35–39
106. Takemura K (2021) Surface plasmon resonance (spr)-and localized spr (lspr)-based virus sensing systems: optical vibration of nano-and micro-metallic materials for the development of next-generation virus detection technology. *Biosensors* 11(8):250

107. Ritchie RH (1957) Plasma losses by fast electrons in thin films. *Phys Rev* 106(5):874–881. <https://doi.org/10.1103/PhysRev.106.874>
108. Johnson PB, Christy RW (1972) Optical constants of the noble metals. *Phys Rev B* 6(12):4370–4379. <https://doi.org/10.1103/PhysRevB.6.4370>
109. Jha R, Singh RK (2023) Tightly focused linearly and radially polarized beam effect on the lspr peak with varying particle size. *Phys Scr* 98(11):115523
110. Belina E, Mankov V, Kisov H, Dimitrova T, Dyankov G (2022) Spectral readout of spr excited in diffraction grating. In: *Journal of Physics: Conference Series*, IOP Publishing, vol 2240, p 012015
111. Raghuvanshi SK, Pandey PS (2022) A numerical study of different metal and prism choices in the surface plasmon resonance biosensor chip for human blood group identification. *IEEE Trans Nanobiosci* 22(2):292–300
112. Kretschmann E, Raether H (1968) Notizen: radiative decay of non radiative surface plasmons excited by light. *Zeitschrift für Naturforschung A* 23(12):2135–2136. <https://doi.org/10.1515/zna-1968-1247>
113. Otto A (1968) Excitation of nonradiative surface plasma waves in silver by the method of frustrated total reflection. *Zeitschrift für Physik A Hadrons and nuclei* 216(4):398–410. <https://doi.org/10.1007/BF01391532>
114. Maier SA (2007) *Plasmonics: fundamentals and applications*. Springer Science & Business Media
115. Grasseschi D, dos Santos D (2020) Nanomateriais plasmônicos: parte i. fundamentos da espectroscopia de nanopartículas e sua relação com o efeito sers. *Química Nova* 43(10):1463–1481. <https://doi.org/10.21577/0100-4042.20170621>
116. Ugwuoke LC, Maňal T, Krüger TP (2020) Localized surface plasmon resonances of simple tunable plasmonic nanostructures. *Plasmonics* 15:189–200
117. Rahaman M, Aslam MA, He L, Madeira TI, Zahn DR (2021) Plasmonic hot electron induced layer dependent anomalous fröhlich interaction in inse. *Commun Phys* 4(1):172
118. Huang H, Lai J, Lu J, Li Z (2021) Performance enhancement of zno ultraviolet detector by localized surface plasmon resonance of al nanoparticles. *Appl Phys A* 127:1–7
119. Kawamura G, Matsuda A (2022) Nanomaterials for localized surface plasmon resonance-related optical functionalities. In: *Progress in Nanophotonics 7*, Springer, pp 37–70
120. Matsko N (2020) Formation of normal surface plasmon modes in small sodium nanoparticles. *Phys Chem Chem Phys* 22(23):13285–13291
121. Zheng J, Liao F, Wu S, Jones G, Chen TY, Fellowes J, Sudmeier T, McPherson IJ, Wilkinson I, Tsang SCE (2019) Efficient non-dissociative activation of dinitrogen to ammonia over lithium-promoted ruthenium nanoparticles at low pressure. *Angew Chem Int Ed* 58(48):17335–17341
122. Catalán-Gómez S, Bran C, Vázquez M, Vázquez L, Pau J, Redondo-Cubero A (2020) Plasmonic coupling in closed-packed ordered gallium nanoparticles. *Sci Rep* 10(1):4187
123. Fonsaca JE, Moreira MP, Farooq S, de Araujo RE, de Matos CJ, Grasseschi D (2023) Surface plasmon resonance platforms for chemical and bio-sensing. 316–353
124. Irfan I, Golovynskyi S, Bosi M, Seravalli L, Yeshchenko OA, Xue B, Dong D, Lin Y, Qiu R, Li B et al (2021) Enhancement of raman scattering and exciton/trion photoluminescence of monolayer and few-layer mos<sub>2</sub> by ag nanoprisms and nanoparticles: shape and size effects. *J Phys Chem C* 125(7):4119–4132
125. Hossain MK (2020) Nanoassembly of gold nanoparticles: an active substrate for size-dependent surface-enhanced Raman scattering. *Spectrochim Acta Part A Mol Biomol Spectrosc* 242
126. Terekhov P, Shamkhi H, Gurvitz E, Baryshnikova K, Evlyukhin A, Shalin A, Karabchevsky A (2019) Broadband forward scattering from dielectric cubic nanoantenna in lossless media. *Opt Express* 27(8):10924–10935
127. Farooq S (2018) Optical properties of metallic nanoparticles and perspectives for biomedical applications. <https://repositorio.ufpe.br/handle/123456789/32614>
128. Wang L, Hasanzadeh Kafshgari M, Meunier M (2020) Optical properties and applications of plasmonic-metal nanoparticles. *Adv Func Mater* 30(51):2005400
129. SS dos Santos P, MMM de Almeida J, Pastoriza-Santos I, CC Coelho L (2021) Advances in plasmonic sensing at the nir—a review. *Sensors* 21(6):2111
130. Kim KY (2012) *Plasmonics: principles and applications*. BoD—Books on Demand, pp 283–312
131. Seo MJ, Kim GW, Tsalu PV, Moon SW, Ha JW (2020) Role of chemical interface damping for tuning chemical enhancement in resonance surface-enhanced Raman scattering of plasmonic gold nanorods. *Nanoscale Horizons* 5(2):345–349
132. Ahmad Mohamed Ali R, Mita D, Espulgar W, Saito M, Nishide M, Takamatsu H, Yoshikawa H, Tamiya E (2019) Single cell analysis of neutrophils nets by microscopic lspr imaging system. *Micromachines* 11(1):52
133. Jakkiriyana N, Durgachalam M, Rajendiren N, Rathinasamy T, Sengeny P (2023) Metal and media refractive index dependent surface plasmon resonance of single and bimetallic core-shell metal nanocomposites. In: *AIP Conference Proceedings*, AIP Publishing, vol 2861
134. Farooq S, Neves WW, Pandoli O, Del Rosso T, de Lima LM, Dutra RF, de Araujo RE (2018b) Engineering a plasmonic sensing platform for *Candida albicans* antigen identification. *J Nanophotonics* 12(3):033003
135. Muldarisnur M, Fridayanti N, Oktorina E, Zeni E, Elvaswer E, Syukri S (2019) Effect of nanoparticle geometry on sensitivity of metal nanoparticle based sensor. In: *IOP Conference Series: Materials Science and Engineering*, IOP Publishing, vol 578, p 012036
136. Mahmood HZ, Jilani A, Farooq S, Javed Y, Jamil Y, Iqbal J, Ullah S, Wageh S (2021) Plasmon-based label-free biosensor using gold nanosphere for dengue detection. *Curr Comput-Aided Drug Des* 11(11):1340
137. Chen H, Kou X, Yang Z, Ni W, Wang J (2008) Shape-and size-dependent refractive index sensitivity of gold nanoparticles. *Langmuir* 24(10):5233–5237
138. Sun Y, Xia Y (2002) Increased sensitivity of surface plasmon resonance of gold nanoshells compared to that of gold solid colloids in response to environmental changes. *Anal Chem* 74(20):5297–5305
139. Underwood S, Mulvaney P (1994) Effect of the solution refractive index on the color of gold colloids. *Langmuir* 10(10):3427–3430
140. Farooq S, Rativa D, de Araujo RE (2019) Optimizing the sensing performance of sio<sub>2</sub>-au nanoshells. *Plasmonics* 14(6):1519–1526
141. Wu J, Li M, Tang H, Su J, He M, Chen G, Guan L, Tian J (2019) Portable paper sensors for the detection of heavy metals based on light transmission-improved quantification of colorimetric assays. *Analyst* 144(21):6382–6390
142. Farooq S, Mahmood HZ, Rativa D, Bouchonneau N, Lins E, Fontana J, de Araujo RE (2018a) Optimizing gold nanorods dimer structure for sensing platform. In: *2018 SBFoton International Optics and Photonics Conference (SBFoton IOPC)*, IEEE, pp 1–4
143. Jung LS, Campbell CT, Chinowsky TM, Mar MN, Yee SS (1998) Quantitative interpretation of the response of surface plasmon resonance sensors to adsorbed films. *Langmuir* 14(19):5636–5648
144. Nehl CL, Liao H, Hafner JH (2006) Optical properties of star-shaped gold nanoparticles. *Nano Lett* 6(4):683–688
145. Khalavka Y, Becker J, Sonnichsen C (2009) Synthesis of rod-shaped gold nanorattles with improved plasmon sensitivity and catalytic activity. *J Am Chem Soc* 131(5):1871–1875

146. Bukasov R, Shumaker-Parry JS (2007) Highly tunable infrared extinction properties of gold nanocrescents. *Nano Lett* 7(5):1113–1118
147. Burgin J, Liu M, Guyot-Sionnest P (2008) Dielectric sensing with deposited gold bipyramids. *The Journal of Physical Chemistry C* 112(49):19279–19282
148. Verma V, Kala D, Gupta S, Kumar H, Kaushal A, Kuča K, Cruz-Martins N, Kumar D (2021) Leptospira interrogans outer membrane protein-based nanohybrid sensor for the diagnosis of leptospirosis. *Sensors* 21(7):2552
149. Sapna K, Sonia J, Shim YB, Arun A, Prasad KS (2022) Au nanoparticle-based disposable electrochemical sensor for detection of leptospirosis in clinical samples. *ACS Applied Nano Materials* 5(9):12454–12463
150. Mancini RS, Sabaine AE, Castro CE, Carnielli JB, Dietze R, de Oliveira VL, Lanfredi AJ, Kubota LT, Mamián-López MB, Alves WA (2022) Development and validation of a sers-based serological test combined with pls-da method for leishmaniasis detection. *ACS Appl Electron Mater* 4(8):3997–4006
151. Martins BR, Barbosa YO, Andrade CM, Pereira LQ, Simão GF, de Oliveira CJ, Correia D, Oliveira RT Jr, da Silva MV, Silva AC et al (2020) Development of an electrochemical immunosensor for specific detection of visceral leishmaniasis using gold-modified screen-printed carbon electrodes. *Biosensors* 10(8):81
152. Sattarahmady N, Movahedpour A, Heli H, Hatam G (2016) Gold nanoparticles-based biosensing of leishmania major kdna genome: visual and spectrophotometric detections. *Sens Actuators, B Chem* 235:723–731
153. Ahmad A, Wei Y, Syed F, Khan S, Khan GM, Tahir K, Khan AU, Raza M, Khan FU, Yuan Q (2016) Isatis tinctoria mediated synthesis of amphotericin b-bound silver nanoparticles with enhanced photoinduced antileishmanial activity: a novel green approach. *J Photochem Photobiol, B* 161:17–24
154. Moradi M, Sattarahmady N, Rahi A, Hatam G, Sorkhabadi SR, Heli H (2016) A label-free, pcr-free and signal-on electrochemical dna biosensor for leishmania major based on gold nanoleaves. *Talanta* 161:48–53
155. Garcia MFds, Andrade CA, de Melo CP, Gomes DS, Silva LG, Dias RV, Balbino VQ, Oliveira MD (2016) Impedimetric sensor for leishmania infantum genome based on gold nanoparticles dispersed in polyaniline matrix. *J Chem Technol Biotechnol* 91(11):2810–2816
156. Nazari-Vanani R, Sattarahmady N, Yadegari H, Delshadi N, Hatam G, Heli H (2018) Electrochemical quantitation of leishmania infantum based on detection of its kdna genome and transduction of non-spherical gold nanoparticles. *Anal Chim Acta* 1041:40–49
157. Jeon W, Lee S, Manjunatha D, Ban C (2013) A colorimetric aptasensor for the diagnosis of malaria based on cationic polymers and gold nanoparticles. *Anal Biochem* 439(1):11–16
158. Della Ventura B, Banchelli M, Funari R, Illiano A, De Angelis M, Taroni P, Amoresano A, Matteini P, Velotta R (2019) Biosensor surface functionalization by a simple photochemical immobilization of antibodies: experimental characterization by mass spectrometry and surface enhanced raman spectroscopy. *Analyst* 144(23):6871–6880
159. Minopoli A, Della Ventura B, Lenyk B, Gentile F, Tanner JA, Offenhäusser A, Mayer D, Velotta R (2020) Ultrasensitive antibody-aptamer plasmonic biosensor for malaria biomarker detection in whole blood. *Nat Commun* 11(1):6134
160. Brangel P, Sobarzo A, Parolo C, Miller BS, Howes PD, Gelkop S, Lutwama JJ, Dye JM, McKendry RA, Lobel L et al (2018) A serological point-of-care test for the detection of igg antibodies against Ebola virus in human survivors. *ACS Nano* 12(1):63–73
161. Zang F, Su Z, Zhou L, Konduru K, Kaplan G, Chou SY (2019) Ultrasensitive Ebola virus antigen sensing via 3d nanoantenna arrays. *Adv Mater* 31(30):1902331
162. Chowdhury AD, Takemura K, Khorish IM, Nasrin F, Tun MMN, Morita K, Park EY (2020b) The detection and identification of dengue virus serotypes with quantum dot and aump regulated localized surface plasmon resonance. *Nanoscale Advances* 2(2):699–709
163. Dutta Chowdhury A, Ganganboina AB, Nasrin F, Takemura K, Ra Doong, Utomo DIS, Lee J, Khorish IM, Park EY (2018) Femtomolar detection of dengue virus dna with serotype identification ability. *Anal Chem* 90(21):12464–12474
164. Camara AR, Gouvêa PM, Dias ACM, Braga AM, Dutra RF, de Araujo RE, Carvalho IC (2013) Dengue immunoassay with an lpr fiber optic sensor. *Opt Express* 21(22):27023–27031
165. Dutra RF, Silva AC, Saade J, Guedes MIF, Cordeiro MT (2018) A carbon ink screen-printed immunoelectrode for dengue virus ns1 protein detection based on photosynthesized amine gold nanoparticles. *J Electron Sens* 1:1–12
166. Nascimento HP, Oliveira MD, de Melo CP, Silva GJ, Cordeiro MT, Andrade CA (2011) An impedimetric biosensor for detection of dengue serotype at picomolar concentration based on gold nanoparticles-polyaniline hybrid composites. *Colloids Surf, B* 86(2):414–419
167. Chen SH, Chuang YC, Lu YC, Lin HC, Yang YL, Lin CS (2009) A method of layer-by-layer gold nanoparticle hybridization in a quartz crystal microbalance dna sensing system used to detect dengue virus. *Nanotechnology* 20(21):215501
168. Tung YT, Wu MF, Wang GJ, Hsieh SL (2014) Nanostructured electrochemical biosensor for th0065 detection of the weak binding between the dengue virus and the clec5a receptor. *Nanomedicine: Nanotechnology, Biology and Medicine* 10(6):1335–1341
169. Luna DM, Avelino KY, Cordeiro MT, Andrade CA, Oliveira MD (2015) Electrochemical immunosensor for dengue virus serotypes based on 4-mercaptobenzoic acid modified gold nanoparticles on self-assembled cysteine monolayers. *Sens Actuators, B Chem* 220:565–572
170. Neves WW, Dutra RF, de Araujo RE, Pandoli O, del Rosso T, Siqueira CG, de Lima LM, Pinheiro J (2015) Development of a localized surface plasmon resonance platform for Candida albicans antigen identification. In: 2015 SBMO/IEEE MTT-S International Microwave and Optoelectronics Conference (IMOC), IEEE, pp 1–4
171. Zhou B, Li G, Wu M, Zhou Z, Cai X, Cai J, Zhou J (2022) Monitoring the adhesion and inhibitory activity of Candida albicans on poly-l-lysine modified gold nano-match head arrays. *Adv Mater Interfaces* 9(16):2102590
172. Cajigas S, Alzate D, Orozco J (2020) Gold nanoparticle/dna-based nanobioconjugate for electrochemical detection of zika virus. *Microchim Acta* 187:1–10
173. Adegoke O, Morita M, Kato T, Ito M, Suzuki T, Park EY (2017) Localized surface plasmon resonance-mediated fluorescence signals in plasmonic nanoparticle-quantum dot hybrids for ultrasensitive zika virus rna detection via hairpin hybridization assays. *Biosens Bioelectron* 94:513–522
174. Takemura K, Adegoke O, Suzuki T, Park EY (2019) A localized surface plasmon resonance-amplified immunofluorescence biosensor for ultrasensitive and rapid detection of nonstructural protein 1 of zika virus. *PLoS ONE* 14(1):e0211517
175. Steinmetz M, Lima D, Viana AG, Fujiwara ST, Pessôa CA, Etto RM, Wohnrath K (2019) A sensitive label-free impedimetric dna biosensor based on silsesquioxane-functionalized gold nanoparticles for zika virus detection. *Biosens Bioelectron* 141:111351
176. Camacho SA, Sobral-Filho RG, Aoki PHB, Constantino CJL, Brolo AG (2018) Zika immunoassay based on surface-enhanced Raman scattering nanoprobos. *ACS Sensors* 3(3):587–594
177. Kaushik A, Yndart A, Kumar S, Jayant RD, Vashist A, Brown AN, Li CZ, Nair M (2018) A sensitive electrochemical immunosensor for label-free detection of zika-virus protein. *Sci Rep* 8(1):9700
178. Santos GS, Andrade CA, Bruscky IS, Wanderley LB, Melo FL, Oliveira MD (2017) Impedimetric nanostructured genosensor for

- detection of schistosomiasis in cerebrospinal fluid and serum samples. *J Pharm Biomed Anal* 137:163–169
179. Santos GS, Caldas RG, Melo FL, Bruscky IS, Silva MA, Wanderley LB, Andrade CA, Oliveira MD (2019) Label-free nanostructured biosensor for schistosoma mansoni detection in complex biological fluids. *Talanta* 204:395–401
  180. Barreto-Duarte B, Araújo-Pereira M, Miguez-Pinto JP, Ferreira IB, Menezes RC, Rosier GL, Vinhaes CL, Maggitti-Bezerril M, Villalva-Serra K, Andrade BB (2022) Grand challenges in major tropical diseases. *Front Trop Dis* 3:1037913
  181. Bharadwaj M, Bengtson M, Golverdingen M, Waling L, Dekker C (2021) Diagnosing point-of-care diagnostics for neglected tropical diseases. *PLoS Negl Trop Dis* 15(6):e0009405
  182. Bennuru S, O'Connell EM, Drame PM, Nutman TB (2018) Mining filarial genomes for diagnostic and therapeutic targets. *Trends Parasitol* 34(1):80–90
  183. Fischer C, Jo WK, Haage V, Moreira-Soto A, de Oliveira Filho EF, Drexler JF (2021) Challenges towards serologic diagnostics of emerging arboviruses. *Clin Microbiol Infect* 27(9):1221–1229
  184. Yang H, Ledesma-Amaro R, Gao H, Ren Y, Deng R (2023b) Crispr-based biosensors for pathogenic biosafety. *Biosensors and Bioelectronics* p 115189
  185. Li Y, Man S, Ye S, Liu G, Ma L (2022) Crispr-cas-based detection for food safety problems: current status, challenges, and opportunities. *Compr Rev Food Sci Food Saf* 21(4):3770–3798
  186. Souto DE, Volpe J, Gonçalves CdC, Ramos CH, Kubota LT (2019c) A brief review on the strategy of developing spr-based biosensors for application to the diagnosis of neglected tropical diseases. *Talanta* 205:120122
  187. Wright WF, Simmer PJ, Carroll KC, Auwaerter PG (2022) Progress report: next-generation sequencing, multiplex polymerase chain reaction, and broad-range molecular assays as diagnostic tools for fever of unknown origin investigations in adults. *Clin Infect Dis* 74(5):924–932
  188. Menon S, Mathew MR, Sam S, Keerthi K, Kumar KG (2020) Recent advances and challenges in electrochemical biosensors for emerging and re-emerging infectious diseases. *J Electroanal Chem* 878:114596
  189. Yang SM, Lv S, Zhang W, Cui Y (2022) Microfluidic point-of-care (poc) devices in early diagnosis: a review of opportunities and challenges. *Sensors* 22(4):1620
  190. Desai AN, Kraemer MU, Bhatia S, Cori A, Nouvellet P, Herringer M, Cohn EL, Carrion M, Brownstein JS, Madoff LC et al (2019) Real-time epidemic forecasting: challenges and opportunities. *Health Security* 17(4):268–275
  191. Doiron B, Mota M, Wells MP, Bower R, Mihai A, Li Y, Cohen LF, Alford NM, Petrov PK, Oulton RF, Maier SA (2019) Quantifying figures of merit for localized surface plasmon resonance applications: a materials survey. *ACS Photonics* 6(2):240–259. <https://doi.org/10.1021/acsp Photonics.8b01369>
  192. Ha M, Kim JH, You M, Li Q, Fan C, Nam JM (2019) Multicomponent plasmonic nanoparticles: from heterostructured nanoparticles to colloidal composite nanostructures. *Chem Rev* 119(24):12208–12278. <https://doi.org/10.1021/acs.chemrev.9b00234>
  193. Mayer KM, Hafner JH (2011) Localized surface plasmon resonance sensors. *Chem Rev* 111(6):3828–57. <https://doi.org/10.1021/cr100313v>
  194. Cortie MB, McDonagh AM (2011) Synthesis and optical properties of hybrid and alloy plasmonic nanoparticles. *Chem Rev* 111(6):3713–35. <https://doi.org/10.1021/cr1002529>
  195. Thanh NTK, Maclean N, Mahiddine S (2014) Mechanisms of nucleation and growth of nanoparticles in solution. *Chem Rev* 114(15):7610–7630. <https://doi.org/10.1021/cr400544s>
  196. Toma HE, Zamarion VM, Toma SH, Araki K (2010) The coordination chemistry at gold nanoparticles. *J Braz Chem Soc* 21(7):1158–1176
  197. Huang CC, Chang HT (2006) Selective gold-nanoparticle-based “turn-on” fluorescent sensors for detection of mercury(II) in aqueous solution. *Anal Chem* 78(24):8332–8
  198. Vianna PG, Grasseschi D, Costa GKB, Carvalho ICS, Domingues SH, Fontana J, de Matos CJS (2016) Graphene oxide/gold nanorod nanocomposite for stable surface-enhanced Raman spectroscopy. *ACS Photonics* 3(6):1027–1035. <https://doi.org/10.1021/acsp Photonics.6b00109>
  199. Su S, Zhang C, Yuwen L, Chao J, Zuo X, Liu X, Song C, Fan C, Wang L (2014) Creating SERS hot spots on MoS<sub>2</sub> nanosheets with in situ grown gold nanoparticles. *ACS Appl Mater Interfaces* 6(21):18735–18741. <https://doi.org/10.1021/am5043092>
  200. Santos EdB, Lima ECNL, de Oliveira CS, Sigoli FA, Mazali IO (2014) Fast detection of paracetamol on a gold nanoparticle–chitosan substrate by SERS. *Anal Methods* 6(11):3564. <https://doi.org/10.1039/c4ay00635f>
  201. Quesada-González D, Merkoçi A (2015) Nanoparticle-based lateral flow biosensors. *Biosens Bioelectron* 73:47–63. <https://doi.org/10.1016/j.bios.2015.05.050>
  202. Grasseschi D, Zamarion VM, Araki K, Toma HE (2010) Surface enhanced Raman scattering spot tests: a new insight on Feigl's analysis using gold nanoparticles. *Anal Chem* 82(22):9146–9149. <https://doi.org/10.1021/ac102238f>
  203. Vianna PG, Grasseschi D, Domingues SH, de Matos CJS (2018) Real-time optofluidic surface-enhanced Raman spectroscopy based on a graphene oxide/gold nanorod nanocomposite. *Opt Express* 26(18):22698. <https://doi.org/10.1364/oe.26.022698>
  204. Grasseschi D, Lima FS, Nakamura M, Toma HE (2015) Hyper-spectral dark-field microscopy of gold nanodisks. *Micron* 69:15–20. <https://doi.org/10.1016/j.micron.2014.10.007>
  205. Yuan Z, Cheng J, Cheng X, He Y, Yeung ES (2012) Highly sensitive DNA hybridization detection with single nanoparticle flash-lamp darkfield microscopy. *Analyst* 137(13):2930. <https://doi.org/10.1039/c2an16171k>
  206. de Pereira MLO, de Souza Paiva R, Vasconcelos TL, Oliveira AG, Oliveira Salles M, Toma HE, Grasseschi D, (2020) Photoinduced electron transfer dynamics of AuNPs and Au@PdNPs supported on graphene oxide probed by dark-field hyperspectral microscopy. *Dalton Trans.* <https://doi.org/10.1039/D0DT01018A>
  207. Mauriz E (2020) Clinical applications of visual plasmonic colorimetric sensing. *Sensors* 20(21):6214
  208. Kim YH, Lee J, Kim YE, Chong CK, Pinchemel Y, Reisdörfer F, Coelho JB, Dias RF, Bae PK, Gusmão ZPM, et al. (2018c) Development of a rapid diagnostic test kit to detect igg/igm antibody against zika virus using monoclonal antibodies to the envelope and non-structural protein 1 of the virus. *Korean J Parasitol* 56(1):61
  209. Shrivastava K, Sahu J, Maji P, Sinha D (2017) Label-free selective detection of ampicillin drug in human urine samples using silver nanoparticles as a colorimetric sensing probe. *New J Chem* 41(14):6685–6692
  210. Versiani AF, Martins EM, Andrade LM, Cox L, Pereira GC, Barbosa-Stancio EF, Nogueira ML, Ladeira LO, da Fonseca FG (2020) Nanosensors based on Ispr are able to serologically differentiate dengue from zika infections. *Sci Rep* 10(1):11302
  211. Martinez-Liu C, Machain-Williams C, Martinez-Acuña N, Lozano-Sepulveda S, Galan-Huerta K, Arellanos-Soto D, Meléndez-Villanueva M, Ávalos-Nolazco D, Pérez-Ibarra K, Galindo-Rodríguez S et al (2022) Development of a rapid gold nanoparticle-based lateral flow immunoassay for the detection of dengue virus. *Biosensors* 12(7):495
  212. Stephen BJ, Suchanti S, Jain D, Dhaliwal H, Sharma V, Kaur R, Mishra R, Singh A (2022) Dna biosensor based detection for neglected tropical disease: moving towards smart diagnosis. *Sens Revw (ahead-of-print)* 42(5):517–525

213. Pirich CL, de Freitas RA, Torresi RM, Picheth GF, Sierakowski MR (2017) Piezoelectric immunochip coated with thin films of bacterial cellulose nanocrystals for dengue detection. *Biosens Bioelectron* 92:47–53
214. Ramos-Jesus J, Pontes-de Carvalho LC, Melo SMB, Alcântara-Neves NM, Dutra RF (2016) A gold nanoparticle piezoelectric immunosensor using a recombinant antigen for detecting leishmania infantum antibodies in canine serum. *Biochem Eng J* 110:43–50
215. Lopez GA, Estevez MC, Soler M, Lechuga LM (2017) Recent advances in nanoplasmonic biosensors: applications and lab-on-a-chip integration. *Nanophotonics* 6(1):123–136. <https://doi.org/10.1515/nanoph-2016-0101>
216. Liu Y, Ling J, Huang CZ (2011) Individually color-coded plasmonic nanoparticles for RGB analysis. *Chem Commun* 47(28):8121–8123
217. Yazdian-Robati R, Hedayati N, Dehghani S, Ramezani M, Alibolandani M, Saeedi M, Abnous K, Taghdisi SM (2021) Application of the catalytic activity of gold nanoparticles for development of optical aptasensors. *Anal Biochem* 629:114307
218. Gracie K, Correa E, Mabbott S, Dougan JA, Graham D, Goodacre R, Faulds K (2014) Simultaneous detection and quantification of three bacterial meningitis pathogens by sers. *Chem Sci* 5(3):1030–1040
219. Lee W, Shaban SM, Pyun DG, Kim DH (2019) Solid-phase colorimetric apta-biosensor for thrombin detection. *Thin Solid Films* 686:137428
220. Kim J, Oh SY, Shukla S, Hong SB, Heo NS, Bajpai VK, Chun HS, Jo CH, Choi BG, Huh YS, et al. (2018b) Heteroassembled gold nanoparticles with sandwich-immunoassay lsrp chip format for rapid and sensitive detection of hepatitis b virus surface antigen (hbsag). *Biosens Bioelectron* 107:118–122
221. Nasrin F, Chowdhury AD, Ganganboina AB, Achadu OJ, Hossain F, Yamazaki M, Park EY (2020) Fluorescent and electrochemical dual-mode detection of chikungunya virus e1 protein using fluorophore-embedded and redox probe-encapsulated liposomes. *Microchim Acta* 187:1–11
222. Farooq S, Vital CV, Tikhonowski G, Popov AA, Klimentov SM, Malagon LA, de Araujo RE, Kabashin AV, Rativa D (2023) Thermo-optical performance of bare laser-synthesized tin nano-fluids for direct absorption solar collector applications. *Sol Energy Mater Sol Cells* 252:112203
223. Al-Gebory L, Menguc MP (2020) A review of optical and radiative properties of nanoparticle suspensions: effects of particle stability, agglomeration, and sedimentation. *J Enhanc Heat Transf* 27(3)
224. Mudalige T, Qu H, Van Haute D, Ansar SM, Paredes A, Ingle T (2019) Characterization of nanomaterials: Tools and challenges. *Nanomaterials for Food Applications* pp 313–353
225. Stodden V, Seiler J, Ma Z (2018) An empirical analysis of journal policy effectiveness for computational reproducibility. *Proc Natl Acad Sci* 115(11):2584–2589
226. Vigneshwaran N, Kathe A, Varadarajan P, Nachane R, Balasubramanya R (2007) Functional finishing of cotton fabrics using silver nanoparticles. *J Nanosci Nanotechnol* 7(6):1893–1897
227. Tolaymat TM, El Badawy AM, Genaidy A, Scheckel KG, Luxton TP, Suidan M (2010) An evidence-based environmental perspective of manufactured silver nanoparticle in syntheses and applications: a systematic review and critical appraisal of peer-reviewed scientific papers. *Sci Total Environ* 408(5):999–1006
228. Karthik L, Kumar G, Keswani T, Bhattacharyya A, Reddy BP, Rao KB (2013) Marine actinobacterial mediated gold nanoparticles synthesis and their antimalarial activity. *Nanomed Nanotechnol Biol Med* 9(7):951–960
229. Marimuthu S, Antonisamy AJ, Malayandi S, Rajendran K, Tsai PC, Pugazhendhi A, Ponnusamy VK (2020) Silver nanoparticles in dye effluent treatment: a review on synthesis, treatment methods, mechanisms, photocatalytic degradation, toxic effects and mitigation of toxicity. *J Photochem Photobiol B* 205:111823
230. Tortella G, Rubilar O, Durán N, Diez M, Martínez M, Parada J, Seabra A (2020) Silver nanoparticles: Toxicity in model organisms as an overview of its hazard for human health and the environment. *J Hazard Mater* 390:121974
231. Sawicki K, Czajka M, Matysiak-Kucharek M, Fal B, Drop B, Męczyńska-Wielgosz S, Sikorska K, Kruszewski M, Kapka-Skrzypczak L (2019) Toxicity of metallic nanoparticles in the central nervous system. *Nanotechnol Rev* 8(1):175–200
232. Fan M, Andrade GF, Brolo AG (2020) A review on recent advances in the applications of surface-enhanced Raman scattering in analytical chemistry. *Anal Chim Acta* 1097:1–29. <https://doi.org/10.1016/j.aca.2019.11.049>
233. dos Santos DP (2020) Statistical analysis of surface-enhanced Raman scattering enhancement distributions. *J Phys Chem C* 124(12):6811–6821. <https://doi.org/10.1021/acs.jpcc.9b11574>

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.