



Short Communication

Antimicrobial photodynamic therapy can be an effective adjuvant for surgical wound healing in cattle



Patrícia Valandro^a, Mayara B. Massuda^a, Elidiane Rusch^a, Daniela B. Birgel^a, Philippe P. L. Pereira^b, Fábio P. Sellera^{b,c}, Martha S. Ribeiro^d, Fabio C. Pogliani^b, Eduardo H. Birgel Junior^{a,*}

^a Department of Veterinary Medicine, School of Animal Science and Food Engineering University of São Paulo, Pirassununga, Brazil

^b Department of Internal Medicine, School of Veterinary Medicine and Animal Science, University of São Paulo, São Paulo, Brazil

^c School of Veterinary Medicine, Metropolitan University of Santos, Santos, Brazil

^d Center for Lasers and Applications, Nuclear, and Energy Research Institute, National Commission for Nuclear Energy, São Paulo, SP, Brazil

ARTICLE INFO

Keywords:

Non-antibiotic approach
Photoinactivation
Rumenostomy
Veterinary medicine
Wound healing

ABSTRACT

Background: Rumenostomy is a useful procedure commonly performed in cattle for medical treatment of domestic ruminants with forestomach diseases. Methylene blue (MB)-mediated antimicrobial photodynamic therapy (APDT) has been broadly investigated to treat infected wounds.

Aim: The aim of the study was to evaluate the effectiveness of MB-mediated APDT (MB-APDT) combined with chlorhexidine and zinc oxide ointment on wound healing process after rumenostomy.

Methods: Fourteen Nelore bulls were subjected to rumenostomy procedure. Animals were randomly divided into MB-APDT (MB associated with a red diode laser performed immediately after surgery and repeated on days 3, 5, 7 and 10) and control groups. Daily care included topical cleaning with chlorhexidine 2% followed by topical zinc oxide ointment. Animals were followed-up until the 28th day.

Results: Wounds presented a better post-surgical profile in MB-APDT group when it was compared with the control group. In MB-APDT group, it was also possible to observe less pain on palpation of wounds borders, less edema and inflammatory exudate. Additionally, animals from MB-APDT group were faster discharged from the cattle care facility.

Conclusion: Our results support the use of MB-APDT for the post-surgical management of rumenostomy. This pilot study ratifies the use of APDT in cattle and also suggests that it could be performed for other surgical procedures as a complementary approach or an alternative for topical administration of antibiotics.

1. Background

Rumenostomy is a surgical procedure performed in cattle to get easy access to ruminal lumen through cannula fixation [1]. It has been commonly used for experimental procedures to evaluate ingestion, digestion, ruminal gas production, and also for rumen transfaunation from a healthy donor animal to treat a diseased animal [1,2]. Wound care is usually performed by cleaning with antiseptic solutions, however infection in surgical site may occur due to perioperative contamination [3,4].

Antimicrobial photodynamic therapy (APDT) has been widely investigated in different areas of medical science [5–10], including veterinary medicine [11]. APDT combines light, a photosensitizing

agent and molecular oxygen to produce oxygen reactive species (ROS) that inactivate microbial cells [5–10]. In this regard, methylene blue (MB)-mediated APDT (MB-APDT) has been in the spotlight to treat topical infected wounds, being considered easy to perform, cost-effective and without causing adverse effects [12,13]. Indeed, MB-APDT has been successfully used to treat superficial infections in companion animals [14], exotic and wild animals [15–17] and food-producing animals [18–20].

2. Aim

The aim of the study was to evaluate the viability of MB-APDT as an adjuvant treatment for wound healing after rumenostomy.

* Corresponding author at: Av. Duque de Caxias Norte, 225 – Campus Fernando Costa - USP, Jardim Elite, Pirassununga, SP, 13635-900, Brazil.

E-mail address: ehbirgel@usp.br (E.H. Birgel Junior).

<https://doi.org/10.1016/j.pdpdt.2020.102168>

Received 14 November 2020; Received in revised form 21 December 2020; Accepted 23 December 2020

Available online 23 January 2021

1572-1000/© 2020 Published by Elsevier B.V.

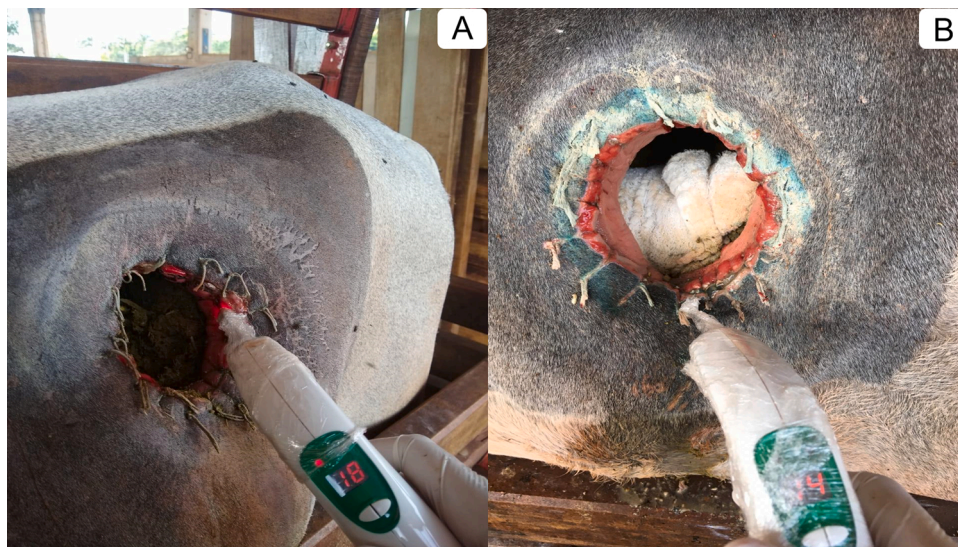


Fig. 1. Representative images of MB-APDT in the space between sutures (rumen-skin junction) (A), and around the wound borders in the postoperative period (B).

Table 1

Clinical evaluation of MB-APDT on the healing of surgical wounds of cattle submitted to ruminostomy during the experimental period. Scores ranged from 0 to 3 (absent, slight, moderate and severe). Values are presented as median and interquartile range. D: Day.

| Clinical sign | | D0 | D3 | D5 | D7 | D10 | D14 | D21 | D28 |
|-------------------|---------|---------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|
| Pain | Control | 0 (0-1) | 2 (1-3) | 2 (0-2) | 1 (1-2) ^A | 0 (0-2) | 0 (0-1) | 0 (0-0) | 0 (0-0) |
| | MB-APDT | 0 (0-1) | 1 (0-1) | 1 (0-1) | 0 (0-1) ^B | 0 (0-0) | 1 (0-1) | 0 (0-1) | 0 (0-0) |
| Edema | Control | 1 (0-1) | 1 (1-3) ^A | 1 (1-3) | 1 (0-3) | 1 (1-2) ^A | 0 (0-2) | 0 (0-1) | 0 (0-0) |
| | MB-APDT | 0 (0-0) | 0 (0-1) ^B | 1 (0-1) | 0 (0-1) | 0 (0-1) ^B | 0 (0-1) | 0 (0-0) | 0 (0-0) |
| Exudate | Control | 0 (0-0) | 1 (0-1) ^A | 1 (1-3) ^A | 1 (0-2) | 1 (1-3) ^A | 2 (1-3) ^A | 1 (1-3) | 1 (0-2) ^A |
| | MB-APDT | 0 (0-0) | 0 (0-0) ^B | 0 (0-0) ^B | 0 (0-1) | 1 (0-1) ^B | 1 (0-1) ^B | 1 (1-2) | 0 (0-0) ^B |
| Suture dehiscence | Control | 0 (0-0) | 0 (0-0) | 0 (0-0) | 1 (0-2) ^A | 3 (1-3) ^A | 3 (1-3) ^A | 2 (0-3) ^A | 0 (0-2) |
| | MB-APDT | 0 (0-0) | 0 (0-0) | 0 (0-0) | 0 (0-1) ^B | 0 (0-1) ^B | 0 (0-1) ^B | 0 (0-0) ^B | 0 (0-0) |

Uppercase letters represent statistically significant differences between groups following Friedman test ($p < 0.05$).

3. Methods

This study was approved by the local animal care and use committee of the School of Animal Science and Food Engineering, University of São Paulo (number: 2368040719/ID001322).

Fourteen 10 months old healthy Nelore bulls, weighing \cong 310 kg were randomly divided in MB-APDT ($n = 7$) and control ($n = 7$) groups, kept in individual stalls, receiving corn silage and water *ad libitum* during the experimental period.

Prior to the surgery, the hair at the left-paralumbar fossa was clipped, followed by local antiseptis and local anesthesia performed with 2% lidocaine infiltrated accordingly the inverted L-block technique [21]. Rumenostomy surgery [1,22] was performed and no interurrences have occurred. Immediately after surgery, penicillin and rifamycin were topically applied, and a silicon cannula was placed through the rumenostomy site. Daily care included topical cleaning with chlorhexidine 2% followed by topical zinc oxide ointment. All sutures were removed at day 12, and animals were followed-up until the 28th day. We also controlled external factors on wound recovery, such as fecal contamination and the influence of environmental humidity since all animals were maintained in the same conditions.

Animals in the MB-APDT group were treated with topical administration of 15 mL of MB aqueous solution at a concentration of 0.01 % (Sigma-Aldrich, USA) directly on the surgical wounds through a syringe. After 5 min, we irradiated punctually the wounds with a diode laser emitting a wavelength of 660 nm, radiant exposure of 70 J/cm², 2 J and 200 s per point (total of 30 points), 100 mW, spot size 0.028 cm² and irradiance of 3.5 W/cm² (TheraVet, DMC®, São Carlos, SP, Brazil) (Fig. 1A-B). The MB-APDT protocol was performed immediately after

surgery and repeated on days 3, 5, 7 and 10. Clinical evaluation was made twice a week during 28 days, and was based on reduction of inflammatory signs (edema and inflammatory exudate), pain assessment (determined by manual palpation of the wound borders and observation of the animals' reactivity), and suture dehiscence. Clinical signs were scored from 0 to 3 (absent, slight, moderate and severe). Data were compared by Friedman test and were considered statistically significant when $p < 0.05$.

4. Results

Table 1 summarizes the scores of clinical signs over the experimental period. Although all wounds from both groups healed uneventfully, some aspects are noteworthy to remark and should be taken in consideration. Observe that on day 7 MB-APDT group showed a statistically lower pain score than the control group. Additionally, on day 10, MB-APDT group exhibited less pronounced exudate and edema. Interestingly, the control group presented a higher score for suture dehiscence on days 7, 10, 14 and 21 than the MB-APDT group.

Overall, in MB-APDT group, no necrotic tissue in the surgical wounds was observed during the evaluated period. On the other hand, in control group, 42.9 % (3/7) of lesions presented tissue necrosis. In the same way, while suture dehiscence occurred in 71.4 % (5/7) of wounds in the control group, only 14.3 % (1/7) was evidenced in MB-APDT group.

The color of the wounds ranged between bright pink, red bright and blackish, however there was no difference between the groups. In brief, in MB-APDT group, it was possible to observe less pain on palpation of wounds borders and animals' reactivity, less edema and inflammatory exudate. Macroscopical assessments of wounds also revealed a better

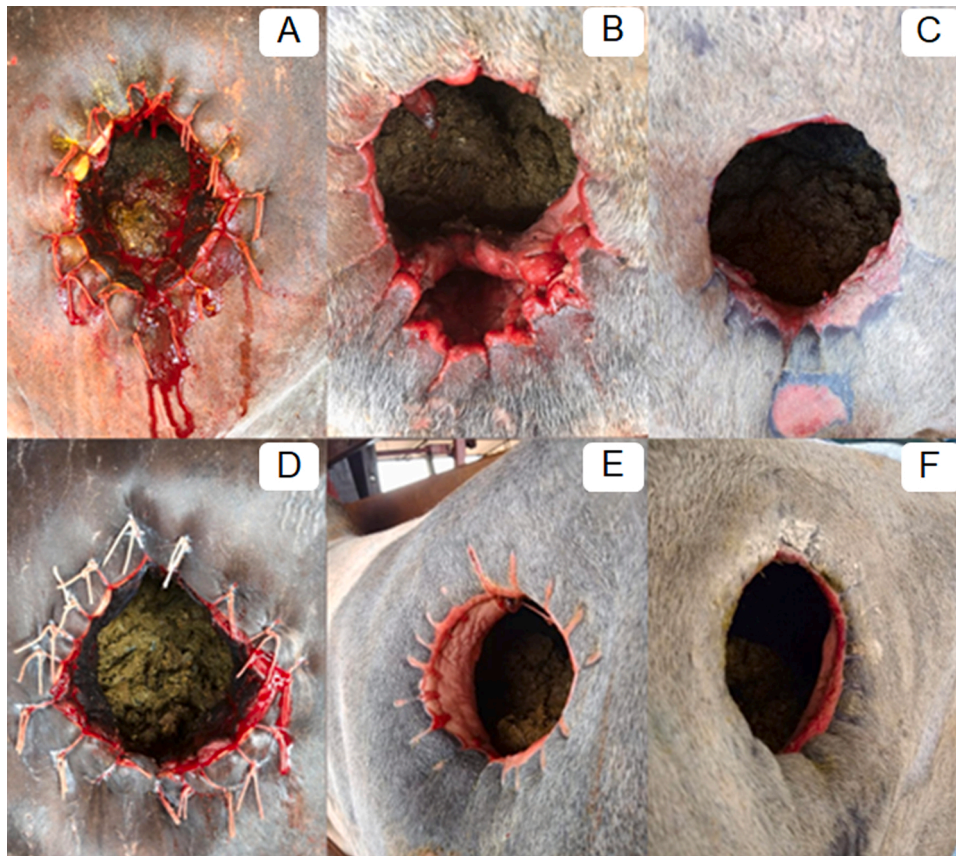


Fig. 2. Representative images of surgical wound healing in cattle. Aspect of wounds from control group immediately after surgical procedure (A), 14 days (B), and 28 days (C). MB-APDT-treated wounds immediately after surgical procedure (D), 14 days (E), and 28 days (F). MB-APDT-treated wounds showed a significant and better improvement at the end of evaluated period.

post-surgical profile in MB-APDT group when it was compared with the control group (Fig. 2A-F).

On the 28th day, 85.7 % (6/7) of animals from MB-APDT group presented complete edge healing, proper cannula fitting, and absence of rumen fluid leakage, being discharged from the cattle care facility. On the other hand, in the control group, only 42.9 % (3/7) showed suitable healing results to be discharged.

5. Discussion

In this study, MB-APDT proved to be a novel and effective adjuvant approach for surgical wound healing in cattle. The choice of MB concentration was based on previous studies that described successful outcomes in Veterinary Medicine and Dentistry [17,23,24]. However, the choice of photosensitizer concentration should consider specific aspects of each wound (bleeding, purulent exudate, etc...). Remarkably, MB-APDT-treated wounds presented better healing than the control group. In this regard, other studies have already demonstrated the ability of MB-APDT to treat food-producing animals with satisfactory results in dermatological and infectious diseases [18–20,25].

In large animal practice, surgical wounds are frequently treated with empirical administration of antibiotics [26] and topical antiseptics [22]. In this regard, a review of outcomes after rumenostomy reported the use of empirical perioperative antibiotics treatment in 92 % of the animals with no further discussion about this strategy regarding the outcomes in the cattle that received elective placement of rumen cannulas [26].

Worryingly, the use of antibiotics in these animals could contribute to the dissemination of antibiotic-resistant bacteria, which is currently considered a critical issue to global public health [27]. Although in our study all animals have received antibiotic therapy, some studies have

reported that MB-APDT could promote a better outcome than antimicrobials [16,28]. Therefore, the replacement of antibiotics for non-antibiotic approaches should be strongly encouraged in contaminated surgical sites and infected wounds such as those from rumenostomy and if there are clinical indications to do so.

In conclusion, we demonstrated that APDT is an effective tool for the post-surgical management of rumenostomy. MB-APDT-treated wounds presented better wound closure, less post-surgery complications, and more animals were discharged from the cattle care facility at the same period. This pilot study ratifies the use of APDT in cattle and also may suggest that it could be performed for other surgical procedures as a complementary approach or an alternative for topical administration of antibiotics. Finally, subsequent studies are essential to clarify the potential applications of APDT in veterinary medicine as well as to assess its feasibility and effectiveness in large animal practice.

Declaration of Competing Interest

The authors report no declarations of interest.

References

- [1] J.W. Lozier, A.J. Niehaus, Surgery of the forestomach, *Vet. Clin. North Am. Food Anim. Pract.* 32 (3) (2016) 617–628, <https://doi.org/10.1016/j.cvfa.2016.05.005>.
- [2] D.L. Harmon, C.J. Richards, Considerations for gastrointestinal cannulations in ruminants, *J. Anim. Sci.* 75 (8) (1997) 2248–2255, <https://doi.org/10.2527/1997.7582248x>.
- [3] A.J. Niehaus, Rumenotomy, *Vet. Clin. North Am. Food Anim. Pract.* 24 (2) (2008) 341–347, <https://doi.org/10.1016/j.cvfa.2008.02.011>.
- [4] M.L. Haven, J.J. Wichtel, D.G. Bristol, J.F. Fetrow, J.W. Spears, Effects of antibiotic prophylaxis on postoperative complications after rumenotomy in cattle, *J. Am. Vet. Med. Assoc.* 200 (9) (1992) 1332–1335.

- [5] M. Wainwright, T. Maisch, S. Nonell, K. Plaetzer, A. Almeida, G.P. Tegós, M. R. Hamblin, Photoantimicrobials-are we afraid of the light? *Lancet Infect. Dis.* 17 (2) (2017) e49–e55, [https://doi.org/10.1016/S1473-3099\(16\)30268-7](https://doi.org/10.1016/S1473-3099(16)30268-7).
- [6] D. Han, Y. Han, J. Li, X. Liu, K.W.K. Yeung, Y. Zheng, Z. Cui, X. Yang, Y. Liang, Z. Li, S. Zhu, X. Yuan, X. Feng, C. Yang, S. Wu, Enhanced photocatalytic activity and photothermal effects of Cu-doped metal-organic frameworks for rapid treatment of bacteria-infected wounds, *Appl. Catal. B* 261 (2020), 118248, <https://doi.org/10.1016/j.apcatb.2019.118248>.
- [7] M. Zhu, X. Liu, L. Tan, Z. Cui, Y. Liang, Z. Li, K.W. Kwok Yeung, S. Wu, Photo-responsive chitosan/Ag/MoS₂ for rapid bacteria-killing, *J. Hazard. Mater.* 383 (2020) 121122, <https://doi.org/10.1016/j.jhazmat.2019.121122>.
- [8] M. Li, L. Li, K. Su, X. Liu, T. Zhang, Y. Liang, D. Jing, X. Yang, D. Zheng, Z. Cui, Z. Li, S. Zhu, K. Yeung, Y. Zheng, X. Wang, S. Wu, Highly effective and noninvasive near-infrared eradication of a *Staphylococcus aureus* biofilm on implants by a photoresponsive coating within 20 min, *Adv. Sci.* 6 (17) (2019) 1900599, <https://doi.org/10.1002/adv.201900599>.
- [9] X. Xie, C. Mao, X. Liu, L. Tan, Z. Cui, X. Yang, S. Zhu, Z. Li, X. Yuan, Y. Zheng, K. Yeung, P.K. Chu, S. Wu, Tuning the bandgap of photo-sensitive polydopamine/Ag₃PO₄/graphene oxide coating for rapid, noninvasive disinfection of implants, *ACS Cent. Sci.* 4 (6) (2018) 724–738, <https://doi.org/10.1021/acscentsci.8b00177>.
- [10] K. Xiong, J. Li, L. Tan, Z. Cui, Z. Li, S. Wu, Y. Liang, S. Zhu, X. Liu, Ag₂S decorated nanocubes with enhanced near-infrared photothermal and photodynamic properties for rapid sterilization, *Colloid Interface Sci. Commun.* 33 (2019) 100201, <https://doi.org/10.1016/j.colcom.2019.100201>.
- [11] F.P. Sellera, C.L. Nascimento, M.S. Ribeiro, *Photodynamic Therapy in Veterinary Medicine: From Basics to Clinical Practice*, Springer, Switzerland, 2017, 228 p.
- [12] J. Oyama, Á.C. Fernandes Herculano Ramos-Milare, D. Lopes Lera-Nonose, V. Nesi-Reis, I. Galhardo Demarchi, S.M. Alessi Aristides, J. Juarez Vieira Teixeira, T. Gomes Verzignassi Silveira, M.V. Campana Lonardoni, Photodynamic therapy in wound healing *in vivo*: a systematic review, *Photodiagnosis Photodyn. Ther.* 30 (2020), 101682, <https://doi.org/10.1016/j.pdpdt.2020.101682>.
- [13] R. Boltes Cecatto, L. Siqueira Magalhães, M. Fernanda Setúbal Destro Rodrigues, C. Pavani, A. Lino-Dos-Santos-Franco, M. Teixeira Gomes, D. Fátima Teixeira Silva, Methylene blue mediated antimicrobial photodynamic therapy in clinical human studies: the state of the art, *Photodiagnosis Photodyn. Ther.* 2020 (2020) 101828, <https://doi.org/10.1016/j.pdpdt.2020.101828>. In press.
- [14] F.P. Sellera, M.R. Fernandes, C.P. Sabino, L.M. de Freitas, L. da Silva, F.C. Pogliani, M.S. Ribeiro, M.R. Hamblin, N. Lincopan, Effective treatment and decolonization of a dog infected with carbapenemase (VIM-2)-producing *Pseudomonas aeruginosa* using probiotic and photodynamic therapies, *Vet. Dermatol.* 30 (2) (2019), <https://doi.org/10.1111/vde.12714>, 170–e52.
- [15] F.P. Sellera, C.P. Sabino, M.S. Ribeiro, L.T. Fernandes, F.C. Pogliani, C.R. Teixeira, G.H. Dutra, C.L. Nascimento, Photodynamic therapy for pododermatitis in penguins, *Zoo Biol.* 33 (4) (2014) 353–356, <https://doi.org/10.1002/zoo.21135>.
- [16] C.L. Nascimento, M.S. Ribeiro, F.P. Sellera, G.H. Dutra, A. Simões, C.R. Teixeira, Comparative study between photodynamic and antibiotic therapies for treatment of footpad dermatitis (bumblefoot) in Magellanic penguins (*Spheniscus magellanicus*), *Photodiagnosis Photodyn. Ther.* 12 (1) (2015) 36–44, <https://doi.org/10.1016/j.pdpdt.2014.12.012>.
- [17] K.F. Grego, M. Carvalho, M. Cunha, T. Knöbl, F.C. Pogliani, J.L. Catão-Dias, S. S. Sant'Anna, M.S. Ribeiro, F.P. Sellera, Antimicrobial photodynamic therapy for infectious stomatitis in snakes: clinical views and microbiological findings, *Photodiagnosis Photodyn. Ther.* 20 (2017) 196–200, <https://doi.org/10.1016/j.pdpdt.2017.10.004>.
- [18] F.P. Sellera, R.G. Gargano, A. Libera, F.J. Benesi, M.R. Azedo, L. de Sá, M. S. Ribeiro, M. da Silva Baptista, F.C. Pogliani, Antimicrobial photodynamic therapy for caseous lymphadenitis abscesses in sheep: report of ten cases, *Photodiagnosis Photodyn. Ther.* 13 (2016) 120–122, <https://doi.org/10.1016/j.pdpdt.2015.12.006>.
- [19] F.P. Sellera, R.G. Gargano, C. Dos Anjos, M. da Silva Baptista, M.S. Ribeiro, F. C. Pogliani, Methylene blue-mediated antimicrobial photodynamic therapy: a novel strategy for digital dermatitis-associated sole ulcer in a cow – a case report, *Photodiagnosis Photodyn. Ther.* 24 (2018) 121–122, <https://doi.org/10.1016/j.pdpdt.2018.09.004>.
- [20] V. Pérez-Laguna, A. Rezusta, J.J. Ramos, L.M. Ferrer, J. Gené, M.J. Revillo, Y. Gilaberte, Daylight photodynamic therapy using methylene blue to treat sheep with dermatophytosis caused by *Arthroderma vanbreusegheemii*, *Small Rumin. Res.* 150 (2017) 97–110, <https://doi.org/10.1016/j.smallrumres.2017.03.011>.
- [21] A. Valverde, M. Sinclair, Ruminant and swine local anesthetic and analgesic techniques, in: K.A. Grimm, L.A. Lamont, W.J. Tranquilli, S.A. Greene, S. A. Robertson (Eds.), *Veterinary Anesthesia and Analgesia*, fifth edition, Blackwell Publishing, 2015, pp. 941–958, <https://doi.org/10.1002/9781119421375.ch51>.
- [22] R.J. Callan, T.J. Applegate, Temporary rumenostomy for the treatment of forestomach diseases and enteral nutrition, *Vet. Clin. North Am. Food Anim. Pract.* 33 (3) (2017) 525–537, <https://doi.org/10.1016/j.cvfa.2017.06.008>.
- [23] V.C. Ribeiro da Silva, F.M. da Motta Silveira, M.G. Barbosa Monteiro, M. da Cruz, A.F. Caldas Júnior, G. Pina Godoy, Photodynamic therapy for treatment of oral mucositis: pilot study with pediatric patients undergoing chemotherapy, *Photodiagnosis Photodyn. Ther.* 21 (2018) 115–120, <https://doi.org/10.1016/j.pdpdt.2017.11.010>.
- [24] G.A. Noro Filho, R.C. Casarin, M.Z. Casati, E.M. Giovani, PDT in non-surgical treatment of periodontitis in HIV patients: a split-mouth, randomized clinical trial, *Lasers Surg. Med.* 44 (4) (2012) 296–302, <https://doi.org/10.1002/lsm.22016>.
- [25] L.H. Moreira, J. de Souza, C.J. de Lima, M. Salgado, A.B. Fernandes, D. Andreani, A.B. Villaverde, R.A. Zângaro, Use of photodynamic therapy in the treatment of bovine subclinical mastitis, *Photodiagnosis Photodyn. Ther.* 21 (2018) 246–251, <https://doi.org/10.1016/j.pdpdt.2017.12.009>.
- [26] A.K. Hartnack, A.J. Niehaus, M. Rousseau, R.L. Pentecost, M.D. Miesner, D. E. Anderson, Indications for and factors relating to outcome after rumenotomy or rumenostomy in cattle: 95 cases (1999–2011), *J. Am. Vet. Med. Assoc.* 247 (2015) 659–664, <https://doi.org/10.2460/javma.247.6.659>.
- [27] EMA (European Medicines Agency) and EFSA (European Food Safety Authority), EMA and EFSA Joint Scientific Opinion on measures to reduce the need to use antimicrobial agents in animal husbandry in the European Union, and the resulting impacts on food safety (RONAFA). [EMA/CVMP/570771/2015], *EFSA J.* 15 (2017) 1–245, <https://doi.org/10.2903/j.efsa.2017.4666>.
- [28] A.R. Scwingel, A.R. Barcessat, S.C. Núñez, M.S. Ribeiro, Antimicrobial photodynamic therapy in the treatment of oral candidiasis in HIV-infected patients, *Photomed. Laser Surg.* 30 (8) (2012) 429–432, <https://doi.org/10.1089/pho.2012.3225>.